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(54) Title: SYSTEM FOR <i>IN VITRO</i> TRANSPOSITION USING MODIFIED TN5 TRANSPOSASE (57) Abstract <p>A system for <i>in vitro</i> transposition includes a donor DNA that includes a transposable element flanked by a pair of bacterial transposon Tn5 outside end repeat sequences, a target DNA into which the transposable element can transpose, and a modified Tn5 transposase having higher binding avidity to the outside end repeat sequences and being less likely to assume an inactive multimer form than wild type Tn5 transposase.</p>		

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5 **SYSTEM FOR *IN VITRO* TRANSPOSITION USING MODIFIED TNS TRANSPOSASE**

CROSS-REFERENCE TO RELATED APPLICATION

 This patent application is a continuation-in-part of a patent application entitled "System for *In Vitro* Transposition," filed March 11, 1997, for which no serial
10 number has yet been accorded. Applicants have petitioned for a filing date of September 9, 1996 to be accorded to the parent application.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

 Not applicable.

15 BACKGROUND OF THE INVENTION

 The present invention relates generally to the field of transposable nucleic acid and, more particularly to production and use of a modified transposase enzyme in a system for introducing genetic changes to nucleic acid.

20 Transposable genetic elements are DNA sequences, found in a wide variety of prokaryotic and eukaryotic organisms, that can move or transpose from one position to another position in a genome. *In vivo*, intra-chromosomal transpositions as well as transpositions between chromosomal and non-chromosomal genetic
25 material are known. In several systems, transposition is known to be under the control of a transposase enzyme that is typically encoded by the transposable element. The genetic structures and transposition mechanisms of various transposable elements are summarized, for example, in "Transposable Genetic
30 Elements" in "The Encyclopedia of Molecular Biology," Kendrew and Lawrence, Eds., Blackwell Science, Ltd., Oxford (1994), incorporated herein by reference.

In vitro transposition systems that utilize the particular transposable elements of bacteriophage Mu and bacterial
35 transposon Tn10 have been described, by the research groups of

5 Kiyoshi Mizuuchi and Nancy Kleckner, respectively.

10 The bacteriophage Mu system was first described by Mizuuchi, K., "In Vitro Transposition of Bacteria Phage Mu: A Biochemical Approach to a Novel Replication Reaction," Cell:785-794 (1983) and Craigie, R. et al., "A Defined System for the DNA Strand-Transfer Reaction at the Initiation of Bacteriophage Mu Transposition: Protein and DNA Substrate Requirements," P.N.A.S. U.S.A. 82:7570-7574 (1985). The DNA donor substrate (mini-Mu) for Mu in vitro reaction normally requires six Mu transposase binding sites (three of about 30 bp at each end) and an enhancer sequence located about 1 kb from the left end. The donor plasmid must be supercoiled. Proteins required are Mu-encoded A and B proteins and host-encoded HU and IHF proteins. Lavoie, B.D, and G. Chaconas, "Transposition of phage Mu DNA," Curr. Topics Microbiol. Immunol. 204:83-99 (1995). The Mu-based system is disfavored for in vitro transposition system applications because the Mu termini are complex and sophisticated and because transposition requires additional proteins above and beyond the transposase.

25 The Tn10 system was described by Morisato, D. and N. Kleckner, "Tn10 Transposition and Circle Formation in vitro," Cell 51:101-111 (1987) and by Benjamin, H. W. and N. Kleckner, "Excision Of Tn10 from the Donor Site During Transposition Occurs By Flush Double-Strand Cleavages at the Transposon Termini," P.N.A.S. U.S.A. 89:4648-4652 (1992). The Tn10 system involves the a supercoiled circular DNA molecule carrying the transposable element (or a linear DNA molecule plus *E. coli* IHF protein). The transposable element is defined by complex 42 bp terminal sequences with IHF binding site adjacent to the inverted repeat. In fact, even longer (81 bp) ends of Tn10 were used in reported experiments. Sakai, J. et al., "Identification and Characterization of Pre-Cleavage Synaptic Complex that is an Early Intermediate in Tn10 transposition," E.M.B.O. J. 14:4374-4383 (1995). In the Tn10 system, chemical treatment of the transposase protein is essential to support active transposition. In addition, the termini of the Tn10 element limit its utility in a generalized in vitro

5 transposition system.

Both the Mu- and Tn10-based *in vitro* transposition systems are further limited in that they are active only on covalently closed circular, supercoiled DNA targets. What is desired is a more broadly applicable *in vitro* transposition system that
10 utilizes shorter, more well defined termini and which is active on target DNA of any structure (linear, relaxed circular, and supercoiled circular DNA).

BRIEF SUMMARY OF THE INVENTION

The present invention is summarized in that an *in vitro*
15 transposition system comprises a preparation of a suitably modified transposase of bacterial transposon Tn5, a donor DNA molecule that includes a transposable element, a target DNA molecule into which the transposable element can transpose, all provided in a suitable reaction buffer.

20 The transposable element of the donor DNA molecule is characterized as a transposable DNA sequence of interest, the DNA sequence of interest being flanked at its 5'- and 3'-ends by short repeat sequences that are acted upon *in trans* by Tn5 transposase.

25 The invention is further summarized in that the suitably modified transposase enzyme comprises two classes of differences from wild type Tn5 transposase, where each class has a separate measurable effect upon the overall transposition activity of the enzyme and where a greater effect is observed
30 when both modifications are present. The suitably modified enzyme both (1) binds to the repeat sequences of the donor DNA with greater avidity than wild type Tn5 transposase ("class (1) mutation") and (2) is less likely than the wild type protein to assume an inactive multimeric form ("class (2) mutation"). A
35 suitably modified Tn5 transposase of the present invention that contains both class (1) and class (2) modifications induces at least about 100-fold ($\pm 10\%$) more transposition than the wild type enzyme, when tested in combination in an *in vivo* conjugation assay as described by Weinreich, M.D., "Evidence
40 that the *cis* Preference of the Tn5 Transposase is Caused by

5 Nonproductive Multimerization," Genes and Development 8:2363-
2374 (1994), incorporated herein by reference. Under optimal
conditions, transposition using the modified transposase may be
higher. A modified transposase containing only a class (1)
10 mutation binds to the repeat sequences with sufficiently
greater avidity than the wild type Tn5 transposase that such a
Tn5 transposase induces about 5- to 50-fold more transposition
than the wild type enzyme, when measured *in vivo*. A modified
transposase containing only a class (2) mutation is
15 sufficiently less likely than the wild type Tn5 transposase to
assume the multimeric form that such a Tn5 transposase also
induces about 5- to 50-fold more transposition than the wild
type enzyme, when measured *in vivo*.

In another aspect, the invention is summarized in that a
method for transposing the transposable element from the donor
20 DNA into the target DNA *in vitro* includes the steps of mixing
together the suitably modified Tn5 transposase protein, the
donor DNA, and the target DNA in a suitable reaction buffer,
allowing the enzyme to bind to the flanking repeat sequences of
the donor DNA at a temperature greater than 0°C, but no higher
25 than about 28°C, and then raising the temperature to
physiological temperature (about 37°C) whereupon cleavage and
strand transfer can occur.

It is an object of the present invention to provide a
useful *in vitro* transposition system having few structural
30 requirements and high efficiency.

It is another object of the present invention to provide a
method that can be broadly applied in various ways, such as to
create absolute defective mutants, to provide selective markers
to target DNA, to provide portable regions of homology to a
35 target DNA, to facilitate insertion of specialized DNA
sequences into target DNA, to provide primer binding sites or
tags for DNA sequencing, to facilitate production of genetic
fusions for gene expression studies and protein domain mapping,
as well as to bring together other desired combinations of DNA
40 sequences (combinatorial genetics).

It is a feature of the present invention that the modified

5 transposase enzyme binds more tightly to DNA than does wild type Tn5 transposase.

It is an advantage of the present invention that the modified transposase facilitates *in vitro* transposition reaction rates of at least about 100-fold higher than can be achieved using wild type transposase (as measured *in vivo*). It is noted that the wild-type Tn5 transposase shows no detectable *in vitro* activity in the system of the present invention. Thus, while it is difficult to calculate an upper limit to the increase in activity, it is clear that hundreds, if not thousands, of colonies are observed when the products of *in vitro* transposition are assayed *in vivo*.

It is another advantage of the present invention that *in vitro* transposition using this system can utilize donor DNA and target DNA that is circular or linear.

It is yet another advantage of the present invention that *in vitro* transposition using this system requires no outside high energy source and no other protein other than the modified transposase.

Other objects, features, and advantages of the present invention will become apparent upon consideration of the following detailed description.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

Fig. 1 depicts test plasmid pRZTL1, used herein to demonstrate transposition *in vitro* of a transposable element located between a pair of Tn5 outside end termini. Plasmid pRZTL1 is also shown and described in SEQ ID NO:3.

Fig. 2 depicts an electrophoretic analysis of plasmid pRZTL1 before and after *in vitro* transposition. Data obtained using both circular and linear plasmid substrates are shown.

Fig. 3 is an electrophoretic analysis of plasmid pRZTL1 after *in vitro* transposition, including further analysis of the molecular species obtained using circular and linear plasmid substrates.

Fig. 4 shows plasmids pRZ1496, pRZ5451 and pRZTL1, which are detailed in the specification.

5 Fig. 5 shows a plot of papillae per colony over time for various mutant OE sequences tested *in vivo* against EK54/MA56 transposase.

Fig. 6 shows a plot of papillae per colony over time for various mutant OE sequences with a smaller Y-axis than is shown
10 in Fig. 5 tested against EK54/MA56 transposase.

Fig. 7 shows a plot of papillae per colony over time for various mutant OE sequences tested against MA56 Tn5 transposase.

Fig. 8 shows *in vivo* transposition using two preferred
15 mutants, tested against MA56 and EK54/MA56 transposase.

DETAILED DESCRIPTION OF THE INVENTION

It will be appreciated that this technique provides a simple, *in vitro* system for introducing any transposable element from a donor DNA into a target DNA. It is generally
20 accepted and understood that Tn5 transposition requires only a pair of OE termini, located to either side of the transposable element. These OE termini are generally thought to be 18 or 19 bases in length and are inverted repeats relative to one another. Johnson, R. C., and W. S. Reznikoff, Nature 304:280
25 (1983), incorporated herein by reference. The Tn5 inverted repeat sequences, which are referred to as "termini" even though they need not be at the termini of the donor DNA molecule, are well known and understood.

Apart from the need to flank the desired transposable element with standard Tn5 outside end ("OE") termini, few other requirements on either the donor DNA or the target DNA are envisioned. It is thought that Tn5 has few, if any, preferences for insertion sites, so it is possible to use the system to introduce desired sequences at random into target
35 DNA. Therefore, it is believed that this method, employing the modified transposase described herein and a simple donor DNA, is broadly applicable to introduce changes into any target DNA, without regard to its nucleotide sequence. It will, thus, be applied to many problems of interest to those skilled in the
40 art of molecular biology.

5 In the method, the modified transposase protein is combined in a suitable reaction buffer with the donor DNA and the target DNA. A suitable reaction buffer permits the transposition reaction to occur. A preferred, but not necessarily optimized, buffer contains spermidine to condense
10 the DNA, glutamate, and magnesium, as well as a detergent, which is preferably 3-[(3-cholamidopropyl) dimethyl-ammonio]-1-propane sulfonate ("CHAPS"). The mixture can be incubated at a temperature greater than 0°C and as high as about 28°C to facilitate binding of the enzyme to the OE termini. Under the
15 buffer conditions used by the inventors in the Examples, a pretreatment temperature of 30°C was not adequate. A preferred temperature range is between 16°C and 28°C. A most preferred pretreatment temperature is about 20°C. Under different buffer conditions, however, it may be possible to use other below-
20 physiological temperatures for the binding step. After a short pretreatment period of time (which has not been optimized, but which may be as little as 30 minutes or as much as 2 hours, and is typically 1 hour), the reaction mixture is diluted with 2 volumes of a suitable reaction buffer and shifted to
25 physiological conditions for several more hours (say 2-3 hours) to permit cleavage and strand transfer to occur. A temperature of 37°C, or thereabouts, is adequate. After about 3 hours, the rate of transposition decreases markedly. The reaction can be stopped by phenol-chloroform extraction and can then be
30 desalted by ethanol precipitation.

When the DNA has been purified using conventional purification tools, it is possible to employ simpler reaction conditions in the *in vitro* transposition method. DNA of sufficiently high purity can be prepared by passing the DNA
35 preparation through a resin of the type now commonly used in the molecular biology laboratory, such as the Qiagen resin of the Qiagen plasmid purification kit (Catalog No. 12162). When such higher quality DNA is employed, CHAPS can be omitted from the reaction buffer. When CHAPS is eliminated from the
40 reaction buffer, the reactants need not be diluted in the manner described above. Also, the low temperature incubation

5 step noted above can be eliminated in favor of a single incubation for cleavage and strand transfer at physiological conditions. A three hour incubation at 37°C is sufficient.

Following the reaction and subsequent extraction steps, transposition can be assayed by introducing the nucleic acid
10 reaction products into suitable bacterial host cells (e.g., *E. coli* K-12 DH5α cells (recA⁻); commercially available from Life Technologies (Gibco-BRL)) preferably by electroporation, described by Dower et al., Nuc. Acids. Res. 16:6127 (1988), and monitoring for evidence of transposition, as is described
15 elsewhere herein.

Those persons skilled in the art will appreciate that apart from the changes noted herein, the transposition reaction can proceed under much the same conditions as would be found in an *in vivo* reaction. Yet, the modified transposase described
20 herein so increases the level of transposition activity that it is now possible to carry out this reaction *in vitro* where this has not previously been possible. The rates of reaction are even greater when the modified transposase is coupled with an optimized buffer and temperature conditions noted herein.

25 In another aspect, the present invention is a preparation of a modified Tn5 transposase enzyme that differs from wild type Tn5 transposase in that it (1) binds to the repeat sequences of the donor DNA with greater avidity than wild type Tn5 transposase and (2) is less likely than the wild type
30 protein to assume an inactive multimeric form. An enzyme having these requirements can be obtained from a bacterial host cell containing an expressible gene for the modified enzyme that is under the control of a promoter active in the host cell. Genetic material that encodes the modified Tn5
35 transposase can be introduced (e.g., by electroporation) into suitable bacterial host cells capable of supporting expression of the genetic material. Known methods for overproducing and preparing other Tn5 transposase mutants are suitably employed. For example, Weinreich, M. D., et al., *supra*, describes a
40 suitable method for overproducing a Tn5 transposase. A second method for purifying Tn5 transposase was described in de la

5 Cruz, N. B., et al., "Characterization of the Tn5 Transposase and Inhibitor Proteins: A Model for the Inhibition of Transposition," J. Bact. 175:6932-6938 (1993), also incorporated herein by reference. It is noted that induction can be carried out at temperatures below 37°C, which is the
10 temperature used by de la Cruz, et al. Temperatures at least in the range of 33 to 37°C are suitable. The inventors have determined that the method for preparing the modified transposase of the present invention is not critical to success of the method, as various preparation strategies have been used
15 with equal success.

Alternatively, the protein can be chemically synthesized, in a manner known to the art, using the amino acid sequence attached hereto as SEQ ID NO:2 as a guide. It is also possible to prepare a genetic construct that encodes the modified
20 protein (and associated transcription and translation signals) by using standard recombinant DNA methods familiar to molecular biologists. The genetic material useful for preparing such constructs can be obtained from existing Tn5 constructs, or can be prepared using known methods for introducing mutations into
25 genetic material (e.g., random mutagenesis PCR or site-directed mutagenesis) or some combination of both methods. The genetic sequence that encodes the protein shown in SEQ ID NO:2 is set forth in SEQ ID NO:1.

The nucleic acid and amino acid sequence of wild type Tn5
30 transposase are known and published. N.C.B.I. Accession Number U00004 L19385, incorporated herein by reference.

In a preferred embodiment, the improved avidity of the modified transposase for the repeat sequences for OE termini (class (1) mutation) can be achieved by providing a lysine
35 residue at amino acid 54, which is glutamic acid in wild type Tn5 transposase. The mutation strongly alters the preference of the transposase for OE termini, as opposed to inside end ("IE") termini. The higher binding of this mutation, known as EK54, to OE termini results in a transposition rate that is
40 about 10-fold higher than is seen with wild type transposase. A similar change at position 54 to valine (mutant EV54) also

5 results in somewhat increased binding/transposition for OE
termini, as does a threonine-to-proline change at position 47
(mutant TP47; about 10-fold higher). It is believed that
other, comparable transposase mutations (in one or more amino
acids) that increase binding avidity for OE termini may also be
10 obtained which would function as well or better in the *in vitro*
assay described herein.

One of ordinary skill will also appreciate that changes to
the nucleotide sequences of the short repeat sequences of the
donor DNA may coordinate with other mutation(s) in or near the
15 binding region of the transposase enzyme to achieve the same
increased binding effect, and the resulting 5- to 50-fold
increase in transposition rate. Thus, while the applicants
have exemplified one case of a mutation that improves binding
of the exemplified transposase, it will be understood that
20 other mutations in the transposase, or in the short repeat
sequences, or in both, will also yield transposases that fall
within the scope and spirit of the present invention. A
suitable method for determining the relative avidity for Tn5 OE
termini has been published by Jilk, R. A., et al., "The
25 Organization of the Outside end of Transposon Tn5," J. Bact.
178:1671-79 (1996).

The transposase of the present invention is also less
likely than the wild type protein to assume an inactive
multimeric form. In the preferred embodiment, that class (2)
30 mutation from wild type can be achieved by modifying amino acid
372 (leucine) of wild type Tn5 transposase to a proline (and,
likewise by modifying the corresponding DNA to encode proline).
This mutation, referred to as LP372, has previously been
characterized as a mutation in the dimerization region of the
35 transposase. Weinreich, et al., *supra*. It was noted by
Weinreich et al. that this mutation at position 372 maps to a
region shown previously to be critical for interaction with an
inhibitor of Tn5 transposition. The inhibitor is a protein
encoded by the same gene that encodes the transposase, but
40 which is truncated at the N-terminal end of the protein,
relative to the transposase. The approach of Weinreich et al.

5 for determining the extent to which multimers are formed is
suitable for determining whether a mutation falls within the
scope of this element.

It is thought that when wild type Tn5 transposase
multimerizes, its activity *in trans* is reduced. Presumably, a
10 mutation in the dimerization region reduces or prevents
multimerization, thereby reducing inhibitory activity and
leading to levels of transposition 5- to 50-fold higher than
are seen with the wild type transposase. The LP372 mutation
achieves about 10-fold higher transposition levels than wild
15 type. Likewise, other mutations (including mutations at a one
or more amino acid) that reduce the ability of the transposase
to multimerize would also function in the same manner as the
single mutation at position 372, and would also be suitable in
a transposase of the present invention. It may also be
20 possible to reduce the ability of a Tn5 transposase to
multimerize without altering the wild type sequence in the so-
called dimerization region, for example by adding into the
system another protein or non-protein agent that blocks the
dimerization site. Alternatively, the dimerization region
25 could be removed entirely from the transposase protein.

As was noted above, the inhibitor protein, encoded in
partially overlapping sequence with the transposase, can
interfere with transposase activity. As such, it is desired
that the amount of inhibitor protein be reduced over the amount
30 observed in wild type *in vivo*. For the present assay, the
transposase is used in purified form, and it may be possible to
separate the transposase from the inhibitor (for example,
according to differences in size) before use. However, it is
also possible to genetically eliminate the possibility of
35 having any contaminating inhibitor protein present by removing
its start codon from the gene that encodes the transposase.

An AUG in the wild type Tn5 transposase gene that encodes
methionine at transposase amino acid 56 is the first codon of
the inhibitor protein. However, it has already been shown that
40 replacement of the methionine at position 56 has no apparent
effect upon the transposase activity, but at the same time

5 prevents translation of the inhibitor protein, thus resulting
in a somewhat higher transposition rate. Weigand, T. W. and W.
S. Reznikoff, "Characterization of Two Hypertransposing Tn5
Mutants," J. Bact. 174:1229-1239 (1992), incorporated herein by
reference. In particular, the present inventors have replaced
10 the methionine with an alanine in the preferred embodiment (and
have replaced the methionine-encoding AUG codon with an
alanine-encoding GCC). A preferred transposase of the present
invention therefore includes an amino acid other than
methionine at amino acid position 56, although this change can
15 be considered merely technically advantageous (since it ensures
the absence of the inhibitor from the *in vitro* system) and not
essential to the invention (since other means can be used to
eliminate the inhibitor protein from the *in vitro* system).

The most preferred transposase amino acid sequence known
20 to the inventors differs from the wild type at amino acid
positions 54, 56, and 372. The mutations at positions 54 and
372 separately contribute approximately a 10-fold increase to
the rate of transposition reaction *in vivo*. When the mutations
are combined using standard recombinant techniques into a
25 single molecule containing both classes of mutations, reaction
rates of at least about 100-fold higher than can be achieved
using wild type transposase are observed when the products of
the *in vitro* system are tested *in vivo*. The mutation at
position 56 does not directly affect the transposase activity.

30 Other mutants from wild type that are contemplated to be
likely to contribute to high transposase activity *in vitro*
include, but are not limited to glutaminic acid-to-lysine at
position 110, and glutamic acid to lysine at position 345.

It is, of course, understood that other changes apart from
35 these noted positions can be made to the modified transposase
(or to a construct encoding the modified transposase) without
adversely affecting the transposase activity. For example, it
is well understood that a construct encoding such a transposase
could include changes in the third position of codons such that
40 the encoded amino acid does not differ from that described
herein. In addition, certain codon changes have little or no

5 functional effect upon the transposition activity of the
encoded protein. Finally, other changes may be introduced
which provide yet higher transposition activity in the encoded
protein. It is also specifically envisioned that combinations
of mutations can be combined to encode a modified transposase
10 having even higher transposition activity than has been
exemplified herein. All of these changes are within the scope
of the present invention. It is noted, however, that a
modified transposase containing the EK110 and EK345 mutations
(both described by Weigand and Reznikoff, *supra*, had lower
15 transposase activity than a transposase containing either
mutation alone.

After the enzyme is prepared and purified, as described
supra, it can be used in the *in vitro* transposition reaction
described above to introduce any desired transposable element
20 from a donor DNA into a target DNA. The donor DNA can be
circular or can be linear. If the donor DNA is linear, it is
preferred that the repeat sequences flanking the transposable
element should not be at the termini of the linear fragment but
should rather include some DNA upstream and downstream from the
25 region flanked by the repeat sequences.

As was noted above, Tn5 transposition requires a pair of
eighteen or nineteen base long termini. The wild type Tn5
outside end (OE) sequence (5'-CTGACTCTTATACACAAGT-3') (SEQ ID
NO: 7) has been described. It has been discovered that a
30 transposase-catalyzed *in vitro* transposition frequency at least
as high as that of wild type OE is achieved if the termini in a
construct include bases ATA at positions 10, 11, and 12,
respectively, as well as the nucleotides in common between wild
type OE and IE (e.g., at positions 1-3, 5-9, 13, 14, 16, and
35 optionally 19). The nucleotides at positions 4, 15, 17, and 18
can correspond to the nucleotides found at those positions in
either wild type OE or wild type IE. It is noted that the
transposition frequency can be enhanced over that of wild type
OE if the nucleotide at position 4 is a T. The importance of
40 these particular bases to transposition frequency has not
previously been identified.

5 It is noted that these changes are not intended to encompass every desirable modification to OE. As is described elsewhere herein, these attributes of acceptable termini modifications were identified by screening mutants having randomized differences between IE and OE termini. While the
10 presence in the termini of certain nucleotides is shown herein to be advantageous, other desirable terminal sequences may yet be obtained by screening a larger array of degenerate mutants that include changes at positions other than those tested herein as well as mutants containing nucleotides not tested in
15 the described screening. In addition, it is clear to one skilled in the art that if a different transposase is used, it may still be possible to select other variant termini, more compatible with that particular transposase.

 Among the mutants shown to be desirable and within the
20 scope of the invention are two hyperactive mutant OE sequences that were identified *in vivo*. Although presented here as single stranded sequences, in fact, the wild type and mutant OE sequences include complementary second strands. The first hyperactive mutant, 5'-CTGTCTCTTATACACATCT-3' (SEQ ID NO: 8),
25 differs from the wild type OE sequence at positions 4, 17, and 18, counting from the 5' end, but retains ATA at positions 10-12. The second, 5'-CTGTCTCTTATACAGATCT-3' (SEQ ID NO: 9), differs from the wild type OE sequence at positions 4, 15, 17, and 18, but also retains ATA at positions 10-12. These two
30 hyperactive mutant OE sequences differ from one another only at position 15, where either G or C is present. OE-like activity (or higher activity) is observed in a mutant sequence when it contains ATA at positions 10, 11 and 12. It may be possible to reduce the length of the OE sequence from 19 to 18 nucleotide
35 pairs with little or no effect.

 When one of the identified hyperactive mutant OE sequences flanks a substrate DNA, the *in vivo* transposition frequency of EK54/MA56 transposase is increased approximately 40-60 fold over the frequency that is observed when wild type OE termini
40 flank the transposable DNA. The EK54/MA56 transposase is already known to have an *in vivo* transposition frequency

5 approximately an 8-10 fold higher than wild type transposase, using wild type OE termini. Tn5 transposase having the EK54/MA56 mutation is known to bind with greater avidity to OE and with lesser avidity to the Tn5 inside ends (IE) than wild type transposase.

10 A suitable mutant terminus in a construct for use in the assays of the present invention is characterized biologically as yielding more papillae per colony in a comparable time, say 68 hours, than is observed in colonies harboring wild type OE in a comparable plasmid. Wild type OE can yield about 100
15 papillae per colony when measured 68 hours after plating in a papillation assay using EK54/MA56 transposase, as is described elsewhere herein. A preferred mutant would yield between about 200 and 3000 papillae per colony, and a more preferred mutant between about 1000 and 3000 papillae per colony, when measured
20 in the same assay and time frame. A most preferred mutant would yield between about 2000 and 3000 papillae per colony when assayed under the same conditions. Papillation levels may be even greater than 3000 per colony, although it is difficult to quantitate at such levels.

25 Transposition frequency is also substantially enhanced in the *in vitro* transposition assay of the present invention when substrate DNA is flanked by a preferred mutant OE sequence and a most preferred mutant transposase (comprising EK54/MA56/LP372 mutations) is used. Under those conditions, essentially all of
30 the substrate DNA is converted into transposition products.

The rate of *in vitro* transposition observed using the hyperactive termini is sufficiently high that, in the experience of the inventors, there is no need to select for transposition events. All colonies selected at random after
35 transformation for further study have shown evidence of transposition events.

This advance can represent a significant savings in time and laboratory effort. For example, it is particularly advantageous to be able to improve *in vitro* transposition
40 frequency by modifying DNA rather than by modifying the transposase because as transposase activity increases in host

5 cells, there is an increased likelihood that cells containing the transposase are killed during growth as a result of aberrant DNA transpositions. In contrast, DNA of interest containing the modified OE termini can be grown in sources completely separate from the transposase, thus not putting the
10 host cells at risk.

Without intending to limit the scope of this aspect of this invention, it is apparent that the tested hyperactive termini do not bind with greater avidity to the transposase than do wild type OE termini. Thus, the higher transposition
15 frequency brought about by the hyperactive termini is not due to enhanced binding to transposase.

The transposable element between the termini can include any desired nucleotide sequence. The length of the transposable element between the termini should be at least
20 about 50 base pairs, although smaller inserts may work. No upper limit to the insert size is known. However, it is known that a donor DNA portion of about 300 nucleotides in length can function well. By way of non-limiting examples, the transposable element can include a coding region that encodes a
25 detectable or selectable protein, with or without associated regulatory elements such as promoter, terminator, or the like.

If the element includes such a detectable or selectable coding region without a promoter, it will be possible to identify and map promoters in the target DNA that are uncovered
30 by transposition of the coding region into a position downstream thereof, followed by analysis of the nucleic acid sequences upstream from the transposition site.

Likewise, the element can include a primer binding site that can be transposed into the target DNA, to facilitate
35 sequencing methods or other methods that rely upon the use of primers distributed throughout the target genetic material. Similarly, the method can be used to introduce a desired restriction enzyme site or polylinker, or a site suitable for another type of recombination, such as a cre-lox, into the
40 target.

The invention can be better understood upon consideration

5 of the following examples which are intended to be exemplary and not limiting on the invention.

EXAMPLES

10 To obtain the transposase modified at position 54, the first third of the coding region from an existing DNA clone that encodes the Tn5 transposase but not the inhibitor protein (MA56) was mutagenized according to known methods and DNA fragments containing the mutagenized portion were cloned to produce a library of plasmid clones containing a full length transposase gene. The clones making up the library were
15 transformed into *E. coli* K-12 strain MDW320 bacteria which were plated and grown into colonies. Transposable elements provided in the bacteria on a separate plasmid contained a defective lacZ gene. The separate plasmid, pOXgen386, was described by Weinreich, M. et al., "A functional analysis of the Tn5
20 Transposase: Identification of Domains Required for DNA Binding and Dimerization," J. Mol. Biol. 241:166-177 (1993), incorporated herein by reference. Colonies having elevated transposase activity were selected by screening for blue (LacZ) spots in white colonies grown in the presence of X-gal. This
25 papillation assay was described by Weinreich, et al. (1993), *supra*. The 5'-most third of Tn5 transposase genes from such colonies were sequenced to determine whether a mutation was responsible for the increase in transposase activity. It was determined that a mutation at position 54 to lysine (K)
30 correlated well with the increase in transposase activity. Plasmid pRZ5412-EK54 contains lysine at position 54 as well as the described alanine at position 56.

35 The fragment containing the LP372 mutation was isolated from pRZ4870 (Weinreich et al (1994)) using restriction enzymes NheI and BglIII, and were ligated into NheI-BglIII cut pRZ5412-EK54 to form a recombinant gene having the mutations at positions 54, 56 and 372, as described herein and shown in SEQ ID NO:1. The gene was tested and shown to have at least about a one hundred fold increase in activity relative to wild type
40 Tn5 transposase. Each of the mutants at positions 54 and 372

5 alone had about a 10-fold increase in transposase activity.

The modified transposase protein encoded by the triple-mutant recombinant gene was transferred into commercial T7 expression vector pET-21D (commercially available from Novagen, Madison, WI) by inserting a BspHI/SalI fragment into NhoI/XhoI
10 fragment of the pET-21D vector. This cloning puts the modified transposase gene under the control of the T7 promoter, rather than the natural promoter of the transposase gene. The gene product was overproduced in BL21(DE3)pLysS bacterial host cells, which do not contain the binding site for the enzyme, by
15 specific induction in a fermentation process after cell growth is complete. (See, Studier, F. W., et al., "Use of T7 RNA Polymerase to Direct Expression of Cloned Genes," Methods Enzymol. 185:60-89 (1990)). The transposase was partially purified using the method of de la Cruz, modified by inducing
20 overproduction at 33 or 37°C. After purification, the enzyme preparation was stored at -70°C in a storage buffer (10% glycerol, 0.7M NaCl, 20 mM Tris-HCl, pH 7.5, 0.1% Triton-X100 and 10 mM CHAPS) until use. This storage buffer is to be considered exemplary and not optimized.

25 A single plasmid (pRZTL1, Fig. 1) was constructed to serve as both donor and target DNA in this Example. The complete sequence of the pRZTL1 plasmid DNA is shown and described in SEQ ID NO:3. Plasmid pRZTL1 contains two Tn5 19 base pair OE termini in inverted orientation to each other. Immediately
30 adjacent to one OE sequence is a gene that would encode tetracycline resistance, but for the lack of an upstream promoter. However, the gene is expressed if the tetracycline resistance gene is placed downstream of a transcribed region (e.g., under the control of the promoter that promotes
35 transcription of the chloramphenicol resistance gene also present on pRZTL1). Thus, the test plasmid pRZTL1 can be assayed *in vivo* after the *in vitro* reaction to confirm that transposition has occurred. The plasmid pRZTL1 also includes an origin of replication in the transposable element, which
40 ensures that all transposition products are plasmids that can replicate after introduction in host cells.

5 The following components were used in typical 20 μ l *in vitro* transposition reactions:

Modified transposase: 2 μ l (approximately 0.1 μ g enzyme/ μ l) in storage buffer (10% glycerol, 0.7M NaCl, 20 mM Tris-HCl, pH 7.5, 0.1% Triton-X100 and 10 mM CHAPS)

10 Donor/Target DNA: 18 μ l (approximately 1-2 μ g) in reaction buffer (at final reaction concentrations of 0.1 M potassium glutamate, 25 mM Tris acetate, pH 7.5, 10 mM Mg²⁺-acetate, 50 μ g/ml BSA, 0.5 mM β -mercaptoethanol, 2 mM spermidine, 100 μ g/ml tRNA).

15 At 20°C, the transposase was combined with pRZTL1 DNA for about 60 minutes. Then, the reaction volume was increased by adding two volumes of reaction buffer and the temperature was raised to 37°C for 2-3 hours whereupon cleavage and strand transfer occurred.

20 Efficient *in vitro* transposition was shown to have occurred by *in vivo* and by *in vitro* methods. *In vivo*, many tetracycline-resistant colonies were observed after transferring the nucleic acid product of the reaction into DH5 α bacterial cells. As noted, tetracycline resistance can only
25 arise in this system if the transposable element is transposed downstream from an active promoter elsewhere on the plasmid. A typical transposition frequency was 0.1% of cells that received plasmid DNA, as determined by counting chloramphenicol
30 resistant colonies. However, this number underestimates the total transposition event frequency because the detection system limits the target to 1/16 of the total.

35 Moreover, *in vitro* electrophoretic (1% agarose) and DNA sequencing analyses of DNA isolated from purified colonies revealed products of true transposition events, including both intramolecular and intermolecular events. Results of typical reactions using circular plasmid pRZTL1 substrates are shown in Lanes 4 & 5. Lane 6 of Fig. 2 shows the results obtained using linear plasmid pRZTL1 substrates.

5 The bands were revealed on 1% agarose gels by staining
with SYBR Green (FMC BioProducts) and were scanned on a
Fluorimager SI (Molecular Dynamics). In Figure 2, lane 1 shows
relaxed circle, linear, and closed circle versions of pRZTL1.
Lanes 2 and 3 show intramolecular and intermolecular
10 transposition products after *in vitro* transposition of pRZTL1,
respectively. The products were purified from electroporated
DH5 α cells and were proven by size and sequence analysis to be
genuine transposition products. Lanes 4 and 5 represent
products of two independent *in vitro* reactions using a mixture
15 of closed and relaxed circular test plasmid substrates. In
lane 6, linear pRZTL1 (XhoI-cut) was the reaction substrate.
Lane 7 includes a BstEII digest of lambda DNA as a molecular
weight standard.

Fig. 3 reproduces lanes 4, 5, and 6 of Fig. 2 and shows an
20 analysis of various products, based upon secondary restriction
digest experiments and re-electroporation and DNA sequencing.
The released donor DNA corresponds to the fragment of pRZTL1
that contains the kanamycin resistance gene between the two OE
sequences, or, in the case of the linear substrate, the OE-XhoI
25 fragment. Intermolecular transposition products can be seen
only as relaxed DNA circles. Intramolecular transposition
products are seen as a ladder, which results from conversion of
the initial superhelicity of the substrate into DNA knots. The
reaction is efficient enough to achieve double transposition
30 events that are a combination of inter- and intramolecular
events.

A preliminary investigation was made into the nature of
the termini involved in a transposition reaction. Wild type
Tn5 OE and IE sequences were compared and an effort was
35 undertaken to randomize the nucleotides at each of the seven
positions of difference. A population of oligonucleotides
degenerate at each position of difference was created. Thus,
individual oligonucleotides in the population randomly included
either the nucleotide of the wild type OE or the wild type IE
40 sequence. In this scheme, 2⁷ (128) distinct oligonucleotides
were synthesized using conventional tools. These

oligonucleotides having sequence characteristics of both OE and IE are referred to herein as OE/IE-like sequences. To avoid nomenclature issues that arise because the oligonucleotides are intermediate between OE and IE wild type sequences, the applicants herein note that selected oligonucleotide sequences are compared to the wild type OE rather than to wild type IE, unless specifically noted. It will be appreciated by one skilled in the art that if IE is selected as the reference point, the differences are identical but are identified differently.

The following depicts the positions (x) that were varied in this mutant production scheme. WT OE is shown also at SEQ ID NO: 7, WT IE at SEQ ID NO: 10.

```

5'-CTGACTCTTATACACAAGT-3' (WT OE)
      x      xxx  x  xx      (positions of difference)
5'-CTGTCTCTTGATCAGATCT-3' (WT IE)

```

In addition to the degenerate OE/IE-like sequences, the 37- base long synthetic oligonucleotides also included terminal *Sph*I and *Kpn*I restriction enzyme recognition and cleavage sites for convenient cloning of the degenerate oligonucleotides into plasmid vectors. Thus, a library of randomized termini was created from population of 2⁷ (128) types of degenerate oligonucleotides.

Fig. 4 shows pRZ1496, the complete sequence of which is presented as SEQ ID NO:11. The following features are noted in the sequence:

<u>Feature</u>	<u>Position</u>
WT OE	94-112
LacZ coding	135-3137
LacY coding	3199-4486
LacA coding	4553-6295
tet ^r coding	6669-9442
transposase coding	10683-12111 (Comp. Strand)
Cassette IE	12184-12225
colE1 sequence	127732-19182

The IE cassette shown in Fig. 4 was excised using *Sph*I and *Kpn*I and was replaced, using standard cleavage and ligation methods, by the synthetic termini cassettes comprising OE/IE-

5 like portions. Between the fixed wild type OE sequence and the
OE/IE-like cloned sequence, plasmid pRZ1496 comprises a gene
whose activity can be detected, namely LacZYA, as well as a
selectable marker gene, tet^r . The LacZ gene is defective in
that it lacks suitable transcription and translation initiation
10 signals. The LacZ gene is transcribed and translated only when
it is transposed into a position downstream from such signals.

The resulting clones were transformed using
electroporation into dam^- , $LacZ^-$ bacterial cells, in this case
JCM101/pOXgen cells which were grown at 37°C in LB medium under
15 standard conditions. A dam^- strain is preferred because dam
methylation can inhibit IE utilization and wild type IE
sequences include two dam methylation sites. A dam^- strain
eliminates dam methylation as a consideration in assessing
transposition activity. The Tet^r cells selected were $LacZ^-$;
20 transposition-activated Lac expression was readily detectable
against a negative background. pOXgen is a non-essential F
factor derivative that need not be provided in the host cells.

In some experiments, the EK54/MA56 transposase was encoded
directly by the transformed pRZ1496 plasmid. In other
25 experiments, the pRZ1496 plasmid was modified by deleting a
unique HindIII/EagI fragment (nucleotides 9112-12083) from the
plasmid (see Fig. 4) to prevent transposase production. In the
latter experiments, the host cells were co-transformed with the
HindIII/EagI-deleted plasmid, termed pRZ5451 (Fig. 4), and with
30 an EK54/MA56 transposase-encoding chloramphenicol-resistant
plasmid. In some experiments, a comparable plasmid encoding a
wild type Tn5 transposase was used for comparison.

Transposition frequency was assessed by a papillation
assay that measured the number of blue spots (Lac producing
35 cells or "papillae") in an otherwise white colony. Transformed
cells were plated (approx. 50 colonies per plate) on Glucose
minimal Miller medium (Miller, J., Experiments in Molecular
Genetics, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY
(1972)) containing 0.3% casamino acids, 5-bromo-4-chloro-3-
40 indolyl- β -D-galactoside (40 μ g/ml) and phenyl- β -D-galactoside
(0.05%). The medium contained tetracycline (15 μ g/ml) and,

5 where needed, chloramphenicol (20 μ g/ml). Colonies that survived the selection were evaluated for transposition frequency *in vivo*. Although colonies exhibiting superior papillation were readily apparent to the naked eye, the number of blue spots per colony were determined over a period of
10 several days (approximately 90 hours post-plating).

To show that the high-papillation phenotype was conferred by the end mutations in the plasmids, colonies were re-streaked if they appeared to have papillation levels higher than was observed when wild type IE was included on the plasmid.

15 Colonies picked from the streaked culture plates were themselves picked and cultured. DNA was obtained and purified from the cultured cells, using standard protocols, and was transformed again into "clean" JCM101/pOXgen cells. Papillation levels were again compared with wild type IE-
20 containing plasmids in the above-noted assays, and consistent results were observed.

To obtain DNA for sequencing of the inserted oligonucleotide, cultures were grown from white portions of 117 hyperpapillating colonies, and DNA was prepared from each
25 colony using standard DNA miniprep methods. The DNA sequence of the OE/IE-like portion of 117 clones was determined (42 from transformations using pRZ1496 as the cloning vehicle; 75 from transformations using pRZ5451 as the cloning vehicle). Only 29 unique mutants were observed. Many mutants were isolated
30 multiple times. All mutants that showed the highest papillation frequencies contain OE-derived bases at positions 10, 11, and 12. When the OE-like bases at these positions were maintained, it was impossible to measure the effect on transposition of other changes, since the papillation level was
35 already extremely high.

One thousand five hundred seventy five colonies were screened as described above. The likelihood that all 128 possible mutant sequences were screened was greater than 95%. Thus, it is unlikely that other termini that contribute to a
40 greater transformation frequency will be obtained using the tested transposase.

TABLE I.
trans papillation level of hybrid end sequences with EK54 Tnp

mutant	position	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	papillation level ^a	# of times isolated ^b
IE		c	t	g	T	c	t	c	t	t	G	A	T	A										
OE					A						A	T	A											
1											A	T	A											
2											A	T	A											
3											A	T	A											
4											A	T	A											
5											A	T	A											
6											A	T	A											
7											A	T	A											
8											A	T	A											
9											A	T	A											
10											A	T	A											
11											A	T	A											
12											A	T	A											
13											A	T	A											
14											A	T	A											
15											A	T	A											
16											A	T	A											
17											A	T	A											
18											A	T	A											
19											A	T	A											
20											A	T	A											
21											A	T	A											

All hybrid end sequences isolated on PRZ5451 that papillate more frequently than wt IE, when the EK54 Tnp is expressed from pFMA187, are listed. ^atrans papillation levels of wt IE, wt OE and hybrid end sequences are classified as follows: VL-very low, L-low, M-medium, and H-high. ^bAlthough mutants 12 and 13 were not found in this experiment, they were found in cis papillation screening (Table II).

TABLE II.
cis papillation level of hybrid end sequences with EK54 Tnp

mutant	position	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	papillation level ^a	# of times isolated ^b
IE		c	t	g	T	c	t	c	t	t	G	A	T	A	T	c	a	G	a	T	C	t	L							
OE		c	t	g	A						A	T	A	A	A	C		A	A	G	G	H								
1											A	T	A	A								H								2
2											A	T	A	A		C														0
3											A	T	A	A				A												1
4											A	T	A	A				A												1
5											A	T	A	A																1
6											A	T	A	A				A												2
7											A	T	A	A				A												3
8											A	T	A	A				A												1
9											A	T	A	A																1
10											A	T	A	A																0
11											A	T	A	A				A												2
12											A	T	A	A																3
13											A	T	A	A				A												1
14											A	T	A	A																0
15											A	T	A	A				A												2
16											A	T	A	A																1
17											A	T	A	A				A												1
18											A	T	A	A																2
19											A	T	A	A				A												2
20											A	T	A	A				A												1
21											A	T	A	A				A												4
22											A	T	A	A																1
23											A	T	A	A				A												1
24											A	T	A	A				A												1
25											A	T	A	A				A												1
26											A	T	A	A				A												1
27											A	T	A	A				A												2

All hybrid end sequences isolated on PRZ1496 that papillate more frequently than wt IE, when the EK54 Tnp is expressed from the same plasmid, are listed. ^acis papillation levels of wt IE, wt OE and hybrid end sequences are classified as follows: L-low, M-medium, MH-medium high, and H-high. Although mutants 2, 10 and 14 were not found in this experiment, they were found in trans papillation screening (Table I).

5 Tables I and II report the qualitative papillation level
of mutant constructs carrying the indicated hybrid end
sequences or the wild type OE or IE end sequences. In the
tables, the sequence at each position of the terminus
corresponds to wild type IE unless otherwise noted. The
10 applicants intend that, while the sequences are presented in
shorthand notation, one of ordinary skill can readily determine
the complete 19 base pair sequence of every presented mutant,
and this specification is to be read to include all such
complete sequences. Table I includes data from trials where
15 the EK54 transposase was provided *in trans*; Table II, from
those trials where the EK54 transposase was provided *in cis*.
Although a transposase provided *in cis* is more active in
absolute terms than a transposase provided *in trans*, the *cis* or
trans source of the transposase does not alter the relative *in*
20 *vivo* transposition frequencies of the tested termini.

 Tables I and II show that every mutant that retains ATA at
positions 10, 11, and 12, respectively, had an activity
comparable to, or higher than, wild type OE, regardless of
whether the wild type OE activity was medium (Table I, *trans*)
25 or high (Table II, *cis*). Moreover, whenever that three-base
sequence in a mutant was not ATA, the mutant exhibited lower
papillation activity than wild type OE. It was also noted that
papillation is at least comparable to, and tends to be
significantly higher than, wild type OE when position 4 is a T.

30 Quantitative analysis of papillation levels was difficult,
beyond the comparative levels shown (very low, low, medium,
medium high, and high). However, one skilled in the art can
readily note the papillation level of OE and can recognize
those colonies having comparable or higher levels. It is
35 helpful to observe the papillae with magnification.

 The number of observed papillae increased over time, as is
shown in Figs 5 - 7 which roughly quantitate the papillation
observed in cells transformed separately with 9 clones
containing either distinct synthetic termini cassettes or wild
40 type OE or IE termini. In these 3 figures, each mutant is
identified by its differences from the wild type IE sequence.

5 Note that, among the tested mutants, only mutant 10A/11T/12A had a higher transposition papillation level than wild type OE. That mutant, which would be called mutant 4/15/17/18 when OE is the reference sequence) was the only mutant of those shown in Figs. 5-7 that retained the nucleotides ATA at positions 10,
10 11, and 12. Figs. 5 (y-axis: 0 - 1500 papillae) and 6 (y-axis: 0 - 250 papillae) show papillation using various mutants plus IE and OE controls and the EK54/MA56 enzyme. Fig. 7 (y-axis: 0 - 250 papillae), shows papillation when the same mutant sequences were tested against the wild type (more properly, MA56) transposase. The 10A/11T/12A mutant (SEQ ID NO: 9) yielded significantly more papillae (approximately 3000) in a shorter time (68 hours) with ED54/MA56 transposase than was observed even after 90 hours with the WT OE (approximately 1500). A single OE-like nucleotide at position 15 on an IE-like background also increased papillation frequency.

In vivo transposition frequency was also quantitated in a tetracycline-resistance assay using two sequences having high levels of hyperpapillation. These sequences were 5'-CTGTCTCTTATACACATCT-3' (SEQ ID NO: 8), which differs from the
25 wild type OE sequence at positions 4, 17, and 18, counting from the 5' end, and 5'-CTGTCTCTTATACAGATCT-3' (SEQ ID NO: 9), which differs from the wild type OE at positions 4, 15, 17, and 18. These sequences are considered the preferred mutant termini in an assay using a transposase that contains EK54/MA56 or a
30 transposase that contains MA56. Each sequence was separately engineered into pRZTL1 in place of the plasmid's two wild type OE sequences. A PCR-amplified fragment containing the desired ends flanking the kanamycin resistance gene was readily cloned into the large HindIII fragment of pRZTL1. The resulting
35 plasmids are identical to pRZTL1 except at the indicated termini. For comparison, pRZTL1 and a derivative of pRZTL1 containing two wild type IE sequences were also tested. In the assay, JCM101/pOXgen cells were co-transformed with a test plasmid (pRZTL1 or derivative) and a high copy number amp^r
40 plasmid that encoded either the EK54/MA56 transposase or wild type (MA56) transposase. The host cells become tetracycline

5 resistant only when a transposition event brings the Tet^r gene into downstream proximity with a suitable transcriptional promoter elsewhere on a plasmid or on the chromosome. The total number of cells that received the test plasmids was determined by counting chloramphenicol resistant, ampicillin
10 resistant colonies. Transposition frequency was calculated by taking the ratio of tet^r/cam^ramp^r colonies. Approximately 40 to 60 fold increase over wild type OE in *in vivo* transposition was observed when using either of the mutant termini and EK54/MA56 transposase. Of the two preferred mutant termini, the one
15 containing mutations at three positions relative to the wild type OE sequence yielded a higher increase.

As is shown in Fig. 8, which plots the tested plasmid against the transposition frequency ($\times 10^{-8}$), little
20 transposition was seen when the test plasmid included two IE termini. Somewhat higher transposition was observed when the test plasmid included two OE termini, particularly when the EK54/MA56 transposase was employed. In striking contrast, the combination of the EK54/MA56 transposase with either of the preferred selected ends (containing OE-like bases only at
25 positions 10, 11, and 12, or positions 10, 11, 12, and 15) yielded a great increase in *in vivo* transposition over wild type OE termini.

The preferred hyperactive mutant terminus having the most preferred synthetic terminus sequence 5'-CTGTCTCTTATACACATCT-3'
30 (SEQ ID NO: 8) was provided in place of both WT OE termini in pRZTL1 (Fig. 4) and was tested in the *in vitro* transposition assay of the present invention using the triple mutant transposase described herein. This mutant terminus was chosen for further *in vitro* analysis because its transposition
35 frequency was higher than for the second preferred synthetic terminus and because it has no dam methylation sites, so dam methylation no longer affects transposition frequency. In contrast the 4/15/17/18 mutant does have a dam methylation site.

40 In a preliminary experiment, CHAPS was eliminated from the reaction, but the pre-incubation step was used. The reaction

5 was pre-incubated for 1 hour at 20°C, then diluted two times,
and then incubated for 3 hours at 37°C. About 0.5 µg of DNA
and 0.4µg of transposase was used. The transposition products
were observed on a gel. With the mutant termini, very little
10 of the initial DNA was observed. Numerous bands representing
primary and secondary transposition reaction products were
observed. The reaction mixtures were transformed into DH5α
cells and were plated on chloramphenicol-, tetracycline-, or
kanamycin-containing plates.

Six hundred forty chloramphenicol-resistant colonies were
15 observed. Although these could represent unreacted plasmid,
all such colonies tested (n=12) were sensitive to kanamycin,
which indicates a loss of donor backbone DNA. All twelve
colonies also included plasmids of varied size; 9 of the 12
were characterized as deletion-inversions, the remaining 3 were
20 simple deletions. Seventy nine tetracycline-resistant
colonies were observed, which indicated an activation of the
tet^r gene by transposition.

Eleven kanamycin resistant colonies were observed. This
indicated a low percentage of remaining plasmids carrying the
25 donor backbone DNA.

In a second, similar test, about 1 µg of plasmid DNA and
0.2 µg transposase were used. In this test, the reaction was
incubated without CHAPS at 37°C for 3 hours without pre-
incubation or dilution. Some initial DNA was observed in the
30 gel after the 3 hour reaction. After overnight incubation,
only transposition products were observed.

The 3 hour reaction products were transformed into DH5α
cells and plated as described. About 50% of the
chloramphenicol resistant colonies were sensitive to kanamycin
35 and were presumably transposition products.

The invention is not intended to be limited to the
foregoing examples, but to encompass all such modifications and
variations as come within the scope of the appended claims.
It is envisioned that, in addition to the uses specifically
40 noted herein, other applications will be apparent to the
skilled molecular biologist. In particular, methods for

5 introducing desired mutations into prokaryotic or eukaryotic
DNA are very desirable. For example, at present it is
difficult to knock out a functional eukaryotic gene by
homologous recombination with an inactive version of the gene
that resides on a plasmid. The difficulty arises from the need
10 to flank the gene on the plasmid with extensive upstream and
downstream sequences. Using this system, however, an
inactivating transposable element containing a selectable
marker gene (e.g., neo) can be introduced *in vitro* into a
plasmid that contains the gene that one desires to inactivate.
15 After transposition, the products can be introduced into
suitable host cells. Using standard selection means, one can
recover only cell colonies that contain a plasmid having the
transposable element. Such plasmids can be screened, for
example by restriction analysis, to recover those that contain
20 a disrupted gene. Such clones can then be introduced directly
into eukaryotic cells for homologous recombination and
selection using the same marker gene.

Also, one can use the system to readily insert a PCR-
amplified DNA fragment into a vector, thus avoiding traditional
25 cloning steps entirely. This can be accomplished by (1)
providing suitable a pair of PCR primers containing OE termini
adjacent to the sequence-specific parts of the primers, (2)
performing standard PCR amplification of a desired nucleic acid
fragment, (3) performing the *in vitro* transposition reaction of
30 the present invention using the double-stranded products of PCR
amplification as the donor DNA.

5

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Reznikoff, William S
Gorysin, Igor Y
Zhou, Hong

(ii) TITLE OF INVENTION: System for In Vitro Transposition

(iii) NUMBER OF SEQUENCES: 11

(iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: Quarles & Brady
(B) STREET: 1 South Pinckney Street
(C) CITY: Madison
(D) STATE: WI
(E) COUNTRY: USA
(F) ZIP: 53703

(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Berson, Bennett J
(B) REGISTRATION NUMBER: 37094
(C) REFERENCE/DOCKET NUMBER: 960296.94142

(ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 608/251-5000
(B) TELEFAX: 608-251-9166

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1534 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "Gene encoding modified Tn5
transposase enzyme"

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 93..1523

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CTGACTCTTA TACACAAGTA GCGTCCTGAA CGGAACCTTT CCCGTTTTCC AGGATCTGAT	60
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Met Ile Thr Ser Ala Leu His	
1 5	

5	CGT GCG GCC GAC TGG GCT AAA TCT GTG TTC TCT TCG GCG GCG CTG GGT Arg Ala Ala Asp Trp Ala Lys Ser Val Phe Ser Ser Ala Ala Leu Gly 10 15 20	161
10	GAT CCT CGC CGT ACT GCC CGC TTG GTT AAC GTC GCC GCC CAA TTG GCA Asp Pro Arg Arg Thr Ala Arg Leu Val Asn Val Ala Ala Gln Leu Ala 25 30 35	209
	AAA TAT TCT GGT AAA TCA ATA ACC ATC TCA TCA GAG GGT AGT AAA GCC Lys Tyr Ser Gly Lys Ser Ile Thr Ile Ser Ser Glu Gly Ser Lys Ala 40 45 50 55	257
15	GCC CAG GAA GGC GCT TAC CGA TTT ATC CGC AAT CCC AAC GTT TCT GCC Ala Gln Glu Gly Ala Tyr Arg Phe Ile Arg Asn Pro Asn Val Ser Ala 60 65 70	305
	GAG GCG ATC AGA AAG GCT GGC GCC ATG CAA ACA GTC AAG TTG GCT CAG Glu Ala Ile Arg Lys Ala Gly Ala Met Gln Thr Val Lys Leu Ala Gln 75 80 85	353
20	GAG TTT CCC GAA CTG CTG GCC ATT GAG GAC ACC ACC TCT TTG AGT TAT Glu Phe Pro Glu Leu Leu Ala Ile Glu Asp Thr Thr Ser Leu Ser Tyr 90 95 100	401
25	CGC CAC CAG GTC GCC GAA GAG CTT GGC AAG CTG GGC TCT ATT CAG GAT Arg His Gln Val Ala Glu Glu Leu Gly Lys Leu Gly Ser Ile Gln Asp 105 110 115	449
	AAA TCC CGC GGA TGG TGG GTT CAC TCC GTT CTC TTG CTC GAG GCC ACC Lys Ser Arg Gly Trp Trp Val His Ser Val Leu Leu Glu Ala Thr 120 125 130 135	497
30	ACA TTC CGC ACC GTA GGA TTA CTG CAT CAG GAG TGG TGG ATG CGC CCG Thr Phe Arg Thr Val Gly Leu Leu His Gln Glu Trp Trp Met Arg Pro 140 145 150	545
	GAT GAC CCT GCC GAT GCG GAT GAA AAG GAG AGT GGC AAA TGG CTG GCA Asp Asp Pro Ala Asp Ala Asp Glu Lys Glu Ser Gly Lys Trp Leu Ala 155 160 165	593
35	GCG GCC GCA ACT AGC CGG TTA CGC ATG GGC AGC ATG ATG AGC AAC GTG Ala Ala Ala Thr Ser Arg Leu Arg Met Gly Ser Met Met Ser Asn Val 170 175 180	641
40	ATT GCG GTC TGT GAC CGC GAA GCC GAT ATT CAT GCT TAT CTG CAG GAC Ile Ala Val Cys Asp Arg Glu Ala Asp Ile His Ala Tyr Leu Gln Asp 185 190 195	689
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45	AAG GAC GTA GAG TCT GGG TTG TAT CTG ATC GAC CAT CTG AAG AAC CAA Lys Asp Val Glu Ser Gly Leu Tyr Leu Ile Asp His Leu Lys Asn Gln 220 225 230	785
	CCG GAG TTG GGT GGC TAT CAG ATC AGC ATT CCG CAA AAG GGC GTG GTG Pro Glu Leu Gly Gly Tyr Gln Ile Ser Ile Pro Gln Lys Gly Val Val 235 240 245	833
50	GAT AAA CGC GGT AAA CGT AAA AAT CGA CCA GCC CGC AAG GCG AGC TTG Asp Lys Arg Gly Lys Arg Lys Asn Arg Pro Ala Arg Lys Ala Ser Leu 250 255 260	881
55	AGC CTG CGC AGT GGG CGC ATC ACG CTA AAA CAG GGG AAT ATC ACG CTC Ser Leu Arg Ser Gly Arg Ile Thr Leu Lys Gln Gly Asn Ile Thr Leu 265 270 275	929

5	AAC GCG GTG CTG GCC GAG GAG ATT AAC CCG CCC AAG GGT GAG ACC CCG Asn Ala Val Leu Ala Glu Ile Asn Pro Pro Lys Gly Glu Thr Pro 280 285 290 295	977
10	TTG AAA TGG TTG TTG CTG ACC GGC GAA CCG GTC GAG TCG CTA GCC CAA Leu Lys Trp Leu Leu Thr Gly Glu Pro Val Glu Ser Leu Ala Gln 300 305 310	1025
	GCC TTG CGC GTC ATC GAC ATT TAT ACC CAT CGC TGG CGG ATC GAG GAG Ala Leu Arg Val Ile Asp Ile Tyr Thr His Arg Trp Arg Ile Glu Glu 315 320 325	1073
15	TTC CAT AAG GCA TGG AAA ACC GGA GCA GGA GCC GAG AGG CAA CGC ATG Phe His Lys Ala Trp Lys Thr Gly Ala Gly Ala Glu Arg Gln Arg Met 330 335 340	1121
	GAG GAG CCG GAT AAT CTG GAG CGG ATG GTC TCG ATC CTC TCG TTT GTT Glu Glu Pro Asp Asn Leu Glu Arg Met Val Ser Ile Leu Ser Phe Val 345 350 355	1169
20	GCG GTC AGG CTG TTA CAG CTC AGA GAA AGC TTC ACG CCG CCG CAA GCA Ala Val Arg Leu Leu Gln Leu Arg Glu Ser Phe Thr Pro Pro Gln Ala 360 365 370 375	1217
25	CTC AGG GCG CAA GGG CTG CTA AAG GAA GCG GAA CAC GTA GAA AGC CAG Leu Arg Ala Gln Gly Leu Leu Lys Glu Ala Glu His Val Glu Ser Gln 380 385 390	1265
	TCC GCA GAA ACG GTG CTG ACC CCG GAT GAA TGT CAG CTA CTG GGC TAT Ser Ala Glu Thr Val Leu Thr Pro Asp Glu Cys Gln Leu Leu Gly Tyr 395 400 405	1313
30	CTG GAC AAG GGA AAA CGC AAG CGC AAA GAG AAA GCA GGT AGC TTG CAG Leu Asp Lys Gly Lys Arg Lys Arg Lys Glu Lys Ala Gly Ser Leu Gln 410 415 420	1361
	TGG GCT TAC ATG GCG ATA GCT AGA CTG GGC GGT TTT ATG GAC AGC AAG Trp Ala Tyr Met Ala Ile Ala Arg Leu Gly Gly Phe Met Asp Ser Lys 425 430 435	1409
35	CGA ACC GGA ATT GCC AGC TGG GGC GCC CTC TGG GAA GGT TGG GAA GCC Arg Thr Gly Ile Ala Ser Trp Gly Ala Leu Trp Glu Gly Trp Glu Ala 440 445 450 455	1457
40	CTG CAA AGT AAA CTG GAT GGC TTT CTT GCC GCC AAG GAT CTG ATG GCG Leu Gln Ser Lys Leu Asp Gly Phe Leu Ala Ala Lys Asp Leu Met Ala 460 465 470	1505
	CAG GGG ATC AAG ATC TGA TCAAGAGACA G Gln Gly Ile Lys Ile *	1534
	475	

(2) INFORMATION FOR SEQ ID NO:2:

- 45 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 477 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

- 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Ile Thr Ser Ala Leu His Arg Ala Ala Asp Trp Ala Lys Ser Val
 1 5 10 15

- 34 -

5 Ser Phe Thr Pro Pro Gln Ala Leu Arg Ala Gln Gly Leu Leu Lys Glu
 370 375 380

Ala Glu His Val Glu Ser Gln Ser Ala Glu Thr Val Leu Thr Pro Asp
 385 390 395 400

10 Glu Cys Gln Leu Leu Gly Tyr Leu Asp Lys Gly Lys Arg Lys Arg Lys
 405 410 415

Glu Lys Ala Gly Ser Leu Gln Trp Ala Tyr Met Ala Ile Ala Arg Leu
 420 425 430

Gly Gly Phe Met Asp Ser Lys Arg Thr Gly Ile Ala Ser Trp Gly Ala
 435 440 445

15 Leu Trp Glu Gly Trp Glu Ala Leu Gln Ser Lys Leu Asp Gly Phe Leu
 450 455 460

Ala Ala Lys Asp Leu Met Ala Gln Gly Ile Lys Ile *

465 470 475

(2) INFORMATION FOR SEQ ID NO:3:

- 20 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 5838 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: circular
- 25 (ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Plasmid DNA"
- (vii) IMMEDIATE SOURCE:
 (B) CLONE: pRZTL1
- 30 (ix) FEATURE:
 (A) NAME/KEY: insertion_seq
 (B) LOCATION: 1..19
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 77..1267
 35 (D) OTHER INFORMATION: /function= "tetracycline resistance"
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: complement (2301..2960)
 (D) OTHER INFORMATION: /function= "chloramphenicol resistance"
- 40 (ix) FEATURE:
 (A) NAME/KEY: insertion_seq
 (B) LOCATION: 4564..4582
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 45 (B) LOCATION: 4715..5530
 (D) OTHER INFORMATION: /function= "kanamycin resistance"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CTGACTCTTA TACACAAGTA AGCTTTAATG CGGTAGTTTA TCACAGTTAA ATTGCTAACG 60

CAGTCAGGCA CCGTGT ATG AAA TCT AAC AAT GCG CTC ATC GTC ATC CTC 109

50 Met Lys Ser Asn Asn Ala Leu Ile Val Ile Leu
 480 485

5	GGC ACC GTC ACC CTG GAT GCT GTA GGC ATA GGC TTG GTT ATG CCG GTA Gly Thr Val Thr Leu Asp Ala Val Gly Ile Gly Leu Val Met Pro Val 490 495 500	157
10	CTG CCG GGC CTC TTG CGG GAT ATC GTC CAT TCC GAC AGC ATC GCC AGT Leu Pro Gly Leu Leu Arg Asp Ile Val His Ser Asp Ser Ile Ala Ser 505 510 515 520	205
	CAC TAT GGC GTG CTG CTA GCG CTA TAT GCG TTG ATG CAA TTT CTA TGC His Tyr Gly Val Leu Leu Ala Leu Tyr Ala Leu Met Gln Phe Leu Cys 525 530 535	253
15	GCA CCC GTT CTC GGA GCA CTG TCC GAC CGC TTT GGC CGC CGC CCA GTC Ala Pro Val Leu Gly Ala Leu Ser Asp Arg Phe Gly Arg Arg Pro Val 540 545 550	301
	CTG CTC GCT TCG CTA CTT GGA GCC ACT ATC GAC TAC GCG ATC ATG GCG Leu Leu Ala Ser Leu Leu Gly Ala Thr Ile Asp Tyr Ala Ile Met Ala 555 560 565	349
20	ACC ACA CCC GTC CTG TGG ATC CTC TAC GCC GGA CGC ATC GTG GCC GGC Thr Thr Pro Val Leu Trp Ile Leu Tyr Ala Gly Arg Ile Val Ala Gly 570 575 580	397
25	ATC ACC GGC GCC ACA GGT GCG GTT GCT GGC GCC TAT ATC GCC GAC ATC Ile Thr Gly Ala Thr Gly Ala Val Ala Gly Ala Tyr Ile Ala Asp Ile 585 590 595 600	445
	ACC GAT GGG GAA GAT CGG GCT CGC CAC TTC GGG CTC ATG AGC GCT TGT Thr Asp Gly Glu Asp Arg Ala Arg His Phe Gly Leu Met Ser Ala Cys 605 610 615	493
30	TTC GGC GTG GGT ATG GTG GCA GGC CCC GTG GCC GGG GGA CTG TTG GGC Phe Gly Val Gly Met Val Ala Gly Pro Val Ala Gly Gly Leu Leu Gly 620 625 630	541
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35	CTC AAC CTA CTA CTG GGC TGC TTC CTA ATG CAG GAG TCG CAT AAG GGA Leu Asn Leu Leu Leu Gly Cys Phe Leu Met Gln Glu Ser His Lys Gly 650 655 660	637
40	GAG CGT CGA CCG ATG CCC TTG AGA GCC TTC AAC CCA GTC AGC TCC TTC Glu Arg Arg Pro Met Pro Leu Arg Ala Phe Asn Pro Val Ser Ser Phe 665 670 675 680	685
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45	TTT ATC ATG CAA CTC GTA GGA CAG GTG CCG GCA GCG CTC TGG GTC ATT Phe Ile Met Gln Leu Val Gly Gln Val Pro Ala Ala Leu Trp Val Ile 700 705 710	781
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50	CTT GCG GTA TTC GGA ATC TTG CAC GCC CTC GCT CAA GCC TTC GTC ACT Leu Ala Val Phe Gly Ile Leu His Ala Leu Ala Gln Ala Phe Val Thr 730 735 740	877
55	GGT CCC GCC ACC AAA CGT TTC GGC GAG AAG CAG GCC ATT ATC GCC GGC Gly Pro Ala Thr Lys Arg Phe Gly Glu Lys Gln Ala Ile Ile Ala Gly 745 750 755 760	925

5	ATG GCG GCC GAC GCG CTG GGC TAC GTC TTG CTG GCG TTC GCG ACG CGA Met Ala Ala Asp Ala Leu Gly Tyr Val Leu Leu Ala Phe Ala Thr Arg	973
	765 770 775	
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	780 785 790	
	GGG ATG CCC GCG TTG CAG GCC ATG CTG TCC AGG CAG GTA GAT GAC GAC Gly Met Pro Ala Leu Gln Ala Met Leu Ser Arg Gln Val Asp Asp Asp	1069
	795 800 805	
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	810 815 820	
	TCG ATC ACT GGA CCG CTG ATC GTC ACG GCG ATT TAT GCC GCC TCG GCG Ser Ile Thr Gly Pro Leu Ile Val Thr Ala Ile Tyr Ala Ala Ser Ala	1165
	825 830 835 840	
20	AGC ACA TGG AAC GGG TTG GCA TGG ATT GTA GGC GCC GCC CTA TAC CTT Ser Thr Trp Asn Gly Leu Ala Trp Ile Val Gly Ala Ala Leu Tyr Leu	1213
	845 850 855	
25	GTC TGC CTC CCC GCG TTG CGT CGC GGT GCA TGG AGC CGG GCC ACC TCG Val Cys Leu Pro Ala Leu Arg Arg Gly Ala Trp Ser Arg Ala Thr Ser	1261
	860 865 870	
	ACC TGA ATGGAAGCCG GCGGCACCTC GCTAACGGAT TCACCACTCC AAGAATTGGA Thr *	1317
30	GCCAATCAAT TCTTGCGGAG AACTGTGAAT GCGCAAACCA ACCCTTGGA GAACATATCC	1377
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	TTGAGAAGCA CACGGTCACA CTGCTTCCGG TAGTCAATAA ACCGGTAAAC CAGCAATAGA	2157
	CATAAGCGGC TATTTAACGA CCCTGCCCTG AACCGACGAC CGGGTCGAAT TTGCTTTCGA	2217
	ATTTCTGCCA TTCATCCGCT TATTATCAAT TATTCAGGCG TAGCACCAGG CGTTTAAGGG	2277
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	AATTCATTAA GCATTCTGCC GACATGGAAG CCATCACAGA CGGCATGATG AACCTGAATC	2397

5	GCCAGCGGCA TCAGCACCTT GTCGCCTTGC GTATAATATT TGCCCATGGT GAAAACGGGG	2457
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	TTGGCTGAGA CGAAAAACAT ATTCTCAATA AACCCCTTTAG GGAAATAGGC CAGGTTTTCA	2577
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	TCACTCCAGA GCGATGAAAA CGTTTCAGTT TGCTCATGGA AAACGGTGTA ACAAGGGTGA	2697
10	ACACTATCCC ATATCACCAG CTCACCGTCT TTCATTGCCA TACGGAATTC CGGATGAGCA	2757
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	ACTGACTGAA ATGCCTCAA ATGTTCTTTA CGATGCCATT GGGATATATC AACGGTG GTA	2937
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	CCAGGATTTA TTTATTCTGC GAAGTGATCT TCCGTCACAG GTATTTATTC GGCGCAAAGT	3177
	GCGTCGGGTG ATGCTGCCAA CTTACTGATT TAGTGTATGA TGGTGTTTTT GAGGTGCTCC	3237
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20	CGGCAAAAGC ACCGCCGGAC ATCAGCGCTA GCGGAGTGTA TACTGGCTTA CTATGTTGGC	3357
	ACTGATGAGG GTGTCAGTGA AGTGCTTCAT GTGGCAGGAG AAAAAAGGCT GCACCGGTGC	3417
	GTCAGCAGAA TATGTGATAC AGGATATATT CCGCTTCCTC GCTCACTGAC TCGCTACGCT	3477
	CGGTGCTTCG ACTGCGGCGA GCGGAAATGG CTTACGAACG GGGCGGAGAT TTCCTGGAAG	3537
	ATGCCAGGAA GATACTTAAC AGGGAAGTGA GAGGGCCGCG GCAAAGCCGT TTTTCCATAG	3597
25	GCTCCGCCCC CCTGACAAGC ATCACGAAAT CTGACGCTCA AATCAGTGGT GGCGAAACCC	3657
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	AGGCGGTTTT TTCGTTTTCA GAGCAAGAGA TTACGCGCAG ACCAAAACGA TCTCAAGAAG	4137
	ATCATCTTAT TAATCAGATA AAATATTTCT AGAGGTGAAC CATCACCTTA ATCAAGTTTT	4197
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	GCTTGACGGG GAAAGCCGGC GAACGTGGCG AGAAAGGAAG GGAAGAAAGC GAAAGGAGCG	4317
	GGCGCTAGGG CGCTGGCAAG TGTAGCGGTC ACGCTGCGCG TAACCACCAC ACCCGCCGCG	4377
	CTTAATGCGC CGCTACAGCG CCATTCGCCA TTCAGGCTGC GCAACTGTTG GGAAGGGCGA	4437

5	TCGGTGCGGG CCTCTTCGCT ATTACGCCAG CTGGCGAAAG GGGGATGTGC TGCAAGGCGA	4497
	TTAAGTTGGG TAACGCCAGG GTTTTCCCAG TCACGACGTT GTAAAACGAC GGCCAGTGCC	4557
	AAGCTTACTT GTGTATAAGA GTCAGTCGAC CTGCAGGGGG GGGGGGGAAA GCCACGTTGT	4617
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10	ACTGTCTGCT TACATAAACA GTAATACAAG GGGTGTT ATG AGC CAT ATT CAA CGG Met Ser His Ile Gln Arg 1 5	4732
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15	TTA TAT GGG TAT AAA TGG GCT CGC GAT AAT GTC GGG CAA TCA GGT GCG Leu Tyr Gly Tyr Lys Trp Ala Arg Asp Asn Val Gly Gln Ser Gly Ala 25 30 35	4828
20	ACA ATC TAT CGA TTG TAT GGG AAG CCC GAT GCG CCA GAG TTG TTT CTG Thr Ile Tyr Arg Leu Tyr Gly Lys Pro Asp Ala Pro Glu Leu Phe Leu 40 45 50	4876
	AAA CAT GGC AAA GGT AGC GTT GCC AAT GAT GTT ACA GAT GAG ATG GTC Lys His Gly Lys Gly Ser Val Ala Asn Asp Val Thr Asp Glu Met Val 55 60 65 70	4924
25	AGA CTA AAC TGG CTG ACG GAA TTT ATG CCT CTT CCG ACC ATC AAG CAT Arg Leu Asn Trp Leu Thr Glu Phe Met Pro Leu Pro Thr Ile Lys His 75 80 85	4972
	TTT ATC CGT ACT CCT GAT GAT GCA TGG TTA CTC ACC ACT GCG ATC CCC Phe Ile Arg Thr Pro Asp Asp Ala Trp Leu Leu Thr Thr Ala Ile Pro 90 95 100	5020
30	GGG AAA ACA GCA TTC CAG GTA TTA GAA GAA TAT CCT GAT TCA GGT GAA Gly Lys Thr Ala Phe Gln Val Leu Glu Glu Tyr Pro Asp Ser Gly Glu 105 110 115	5068
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5 ATC CTA TGG AAC TGC CTC GGT GAG TTT TCT CCT TCA TTA CAG AAA CGG 5452
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 235 240 245

CTT TTT CAA AAA TAT GGT ATT GAT AAT CCT GAT ATG AAT AAA TTG CAG 5500
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10 TTT CAT TTG ATG CTC GAT GAG TTT TTC TAA TCAGAATTGG TTAATTGGTT 5550
 Phe His Leu Met Leu Asp Glu Phe Phe *
 265 270

GTAACACTGG CAGAGCATT A CGCTGACTTG ACGGGACGGC GGCTTTGTTG AATAAATCGA 5610

15 ACTTTTGCTG AGTTGAAGGA TCAGATCACG CATCTTCCCG ACAACGCAGA CCGTTCCGTG 5670

GCAAAGCAAA AGTTCAAAAT CACCAACTGG TCCACCTACA ACAAAGCTCT CATCAACCGT 5730

GGCTCCCTCA CTTTCTGGCT GGATGATGGG GCGATTGAGG CCTGGTATGA GTCAGCAACA 5790

CCTTCTTCAC GAGGCAGACC TCAGCGCCCC CCCCCCCTG CAGGTCGA 5838

(2) INFORMATION FOR SEQ ID NO:4:

20 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 397 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Lys Ser Asn Asn Ala Leu Ile Val Ile Leu Gly Thr Val Thr Leu
 1 5 10 15

Asp Ala Val Gly Ile Gly Leu Val Met Pro Val Leu Pro Gly Leu Leu
 20 25 30

30 Arg Asp Ile Val His Ser Asp Ser Ile Ala Ser His Tyr Gly Val Leu
 35 40 45

Leu Ala Leu Tyr Ala Leu Met Gln Phe Leu Cys Ala Pro Val Leu Gly
 50 55 60

35 Ala Leu Ser Asp Arg Phe Gly Arg Arg Pro Val Leu Leu Ala Ser Leu
 65 70 75 80

Leu Gly Ala Thr Ile Asp Tyr Ala Ile Met Ala Thr Thr Pro Val Leu
 85 90 95

Trp Ile Leu Tyr Ala Gly Arg Ile Val Ala Gly Ile Thr Gly Ala Thr
 100 105 110

40 Gly Ala Val Ala Gly Ala Tyr Ile Ala Asp Ile Thr Asp Gly Glu Asp
 115 120 125

Arg Ala Arg His Phe Gly Leu Met Ser Ala Cys Phe Gly Val Gly Met
 130 135 140

45 Val Ala Gly Pro Val Ala Gly Gly Leu Leu Gly Ala Ile Ser Leu His
 145 150 155 160

Ala Pro Phe Leu Ala Ala Ala Val Leu Asn Gly Leu Asn Leu Leu Leu
 165 170 175

5 Gly Cys Phe Leu Met Gln Glu Ser His Lys Gly Glu Arg Arg Pro Met
180 185 190

Pro Leu Arg Ala Phe Asn Pro Val Ser Ser Phe Arg Trp Ala Arg Gly
195 200 205

10 Met Thr Ile Val Ala Ala Leu Met Thr Val Phe Phe Ile Met Gln Leu
210 215 220

Val Gly Gln Val Pro Ala Ala Leu Trp Val Ile Phe Gly Glu Asp Arg
225 230 235 240

Phe Arg Trp Ser Ala Thr Met Ile Gly Leu Ser Leu Ala Val Phe Gly
245 250 255

15 Ile Leu His Ala Leu Ala Gln Ala Phe Val Thr Gly Pro Ala Thr Lys
260 265 270

Arg Phe Gly Glu Lys Gln Ala Ile Ile Ala Gly Met Ala Ala Asp Ala
275 280 285

20 Leu Gly Tyr Val Leu Leu Ala Phe Ala Thr Arg Gly Trp Met Ala Phe
290 295 300

Pro Ile Met Ile Leu Leu Ala Ser Gly Gly Ile Gly Met Pro Ala Leu
305 310 315 320

Gln Ala Met Leu Ser Arg Gln Val Asp Asp Asp His Gln Gly Gln Leu
325 330 335

25 Gln Gly Ser Leu Ala Ala Leu Thr Ser Leu Thr Ser Ile Thr Gly Pro
340 345 350

Leu Ile Val Thr Ala Ile Tyr Ala Ala Ser Ala Ser Thr Trp Asn Gly
355 360 365

30 Leu Ala Trp Ile Val Gly Ala Ala Leu Tyr Leu Val Cys Leu Pro Ala
370 375 380

Leu Arg Arg Gly Ala Trp Ser Arg Ala Thr Ser Thr *

385 390 395

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

35 (A) LENGTH: 220 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

40 Met Glu Lys Lys Ile Thr Gly Tyr Thr Thr Val Asp Ile Ser Gln Trp
1 5 10 15

His Arg Lys Glu His Phe Glu Ala Phe Gln Ser Val Ala Gln Cys Thr
20 25 30

45 Tyr Asn Gln Thr Val Gln Leu Asp Ile Thr Ala Phe Leu Lys Thr Val
35 40 45

Lys Lys Asn Lys His Lys Phe Tyr Pro Ala Phe Ile His Ile Leu Ala
50 55 60

Arg Leu Met Asn Ala His Pro Glu Phe Arg Met Ala Met Lys Asp Gly
65 70 75 80

5 Glu Leu Val Ile Trp Asp Ser Val His Pro Cys Tyr Thr Val Phe His
 85 90 95
 Glu Gln Thr Glu Thr Phe Ser Ser Leu Trp Ser Glu Tyr His Asp Asp
 100 105 110
 10 Phe Arg Gln Phe Leu His Ile Tyr Ser Gln Asp Val Ala Cys Tyr Gly
 115 120 125
 Glu Asn Leu Ala Tyr Phe Pro Lys Gly Phe Ile Glu Asn Met Phe Phe
 130 135 140
 Val Ser Ala Asn Pro Trp Val Ser Phe Thr Ser Phe Asp Leu Asn Val
 145 150 155 160
 15 Ala Asn Met Asp Asn Phe Phe Ala Pro Val Phe Thr Met Gly Lys Tyr
 165 170
 Tyr Thr Gln Gly Asp Lys Val Leu Met Pro Leu Ala Ile Gln Val His
 180 185 190
 20 His Ala Val Cys Asp Gly Phe His Val Gly Arg Met Leu Asn Glu Leu
 195 200 205
 Gln Gln Tyr Cys Asp Glu Trp Gln Gly Gly Ala *
 210 215 220

(2) INFORMATION FOR SEQ ID NO:6:

25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 272 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

30 Met Ser His Ile Gln Arg Glu Thr Ser Cys Ser Arg Pro Arg Leu Asn
 1 5 10 15
 Ser Asn Met Asp Ala Asp Leu Tyr Gly Tyr Lys Trp Ala Arg Asp Asn
 20 25 30
 35 Val Gly Gln Ser Gly Ala Thr Ile Tyr Arg Leu Tyr Gly Lys Pro Asp
 35 40 45
 Ala Pro Glu Leu Phe Leu Lys His Gly Lys Gly Ser Val Ala Asn Asp
 50 55 60
 Val Thr Asp Glu Met Val Arg Leu Asn Trp Leu Thr Glu Phe Met Pro
 65 70 75 80
 40 Leu Pro Thr Ile Lys His Phe Ile Arg Thr Pro Asp Asp Ala Trp Leu
 85 90 95
 Leu Thr Thr Ala Ile Pro Gly Lys Thr Ala Phe Gln Val Leu Glu Glu
 100 105 110
 45 Tyr Pro Asp Ser Gly Glu Asn Ile Val Asp Ala Leu Ala Val Phe Leu
 115 120 125
 Arg Arg Leu His Ser Ile Pro Val Cys Asn Cys Pro Phe Asn Ser Asp
 130 135 140
 Arg Val Phe Arg Leu Ala Gln Ala Gln Ser Arg Met Asn Asn Gly Leu
 145 150 155 160

5 Val Asp Ala Ser Asp Phe Asp Asp Glu Arg Asn Gly Trp Pro Val Glu
165 170 175
Gln Val Trp Lys Glu Met His Lys Leu Leu Pro Phe Ser Pro Asp Ser
180 185 190
10 Val Val Thr His Gly Asp Phe Ser Leu Asp Asn Leu Ile Phe Asp Glu
195 200 205
Gly Lys Leu Ile Gly Cys Ile Asp Val Gly Arg Val Gly Ile Ala Asp
210 215 220
Arg Tyr Gln Asp Leu Ala Ile Leu Trp Asn Cys Leu Gly Glu Phe Ser
225 230 235 240
15 Pro Ser Leu Gln Lys Arg Leu Phe Gln Lys Tyr Gly Ile Asp Asn Pro
245 250 255
Asp Met Asn Lys Leu Gln Phe His Leu Met Leu Asp Glu Phe Phe *
260 265 270

(2) INFORMATION FOR SEQ ID NO:7:

- 20 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- 25 (ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "Tn5 wild type outside end"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CTGACTCTTA TACACAAGT

19

(2) INFORMATION FOR SEQ ID NO:8:

- 30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- 35 (ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "Tn5 mutant outside end"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

CTGTCTCTTA TACACATCT

19

(2) INFORMATION FOR SEQ ID NO:9:

- 40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
- 45 (ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "Tn5 mutant outside end"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CTGTCTCTTA TACAGATCT

19

5 (2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 10 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Tn5 wild type inside end"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
 CTGTCTCTTG ATCAGATCT 19

15 (2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19182 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 20 (D) TOPOLOGY: circular

(ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Plasmid pRZ4196"

(ix) FEATURE:
 (A) NAME/KEY: repeat_unit
 (B) LOCATION: 94..112
 (D) OTHER INFORMATION: /note= "Wild type OE sequence"

25 (ix) FEATURE:
 (A) NAME/KEY: repeat_unit
 (B) LOCATION: 12184..12225
 (D) OTHER INFORMATION: /note= "Cassette IE"

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TTCCTGTAAC AATAGCAATA CCCCAAATAC CTAATGTAGT TCCAGCAAGC AAGCTAAAAA	60
GTAAAGCAAC AACATAACTC ACCCCTGCAT CTGCTGACTC TTATACACAA GTAGCGTCCC	120
GGGATCGGGA TCCCGTCGTT TTACAACGTC GTGACTGGGA AAACCCTGGC GTTACCCAAC	180
35 TTAATCGCCT TGCAGCACAT CCCCCTTTTCG CCAGCTGGCG TAATAGCGAA GAGGCCCGCA	240
CCGATCGCCC TTCCAACAG TTGCGCAGCC TGAATGGCGA ATGGCGCTTT GCCTGGTTTC	300
CGGCACCAGA AGCGGTGCCG GAAAGCTGGC TGGAGTGCGA TCTTCCTGAG GCCGATACTG	360
TCGTCGTCCC CTCAAACCTGG CAGATGCACG GTTACGATGC GCCCATCTAC ACCAACGTAA	420
CCTATCCCAT TACGGTCAAT CCGCCGTTTG TTCCACGGA GAATCCGACG GGTTGTTACT	480
40 CGCTCACATT TAATGTTGAT GAAAGCTGGC TACAGGAAGG CCAGACGCGA ATTATTTTTC	540
ATGGCGTTAA CTCGGCGTTT CATCTGTGGT GCAACGGGCG CTGGGTCGGT TACGGCCAGG	600
ACAGTCGTTT GCCGTCTGAA TTTGACCTGA GCGCATTTTT ACGCGCCGGA GAAAACCGCC	660
TCGCGGTGAT GGTGCTGCGT TGGAGTGACG GCAGTTATCT GGAAGATCAG GATATGTGGC	720
GGATGAGCGG CATTTTCCGT GACGTCTCGT TGCTGCATAA ACCGACTACA CAAATCAGCG	780
45 ATTTCCATGT TGCCACTCGC TTTAATGATG ATTTTCAGCCG CGCTGTACTG GAGGCTGAAG	840
TTCAGATGTG CGGCGAGTTG CGTGACTACC TACGGGTAAC AGTTTCTTTA TGGCAGGGTG	900

5	AAACGCAGGT CGCCAGCGGC ACCGCGCCTT TCGGCGGTGA AATTATCGAT GAGCGTGGTG	960
	GTTATGCCGA TCGCGTCACA CTACGTCTGA ACGTCGAAAA CCCGAAACTG TGGAGCGCCG	1020
	AAATCCCGAA TCTCTATCGT GCGGTGGTTG AACTGCACAC CGCCGACGGC ACGCTGATTG	1080
	AAGCAGAAGC CTGCGATGTC GGTTCCTCGG AGGTGCGGAT TGAAAATGGT CTGCTGCTGC	1140
	TGAACGGCAA GCCGTTGCTG ATTTCGAGGCG TTAACCGTCA CGAGCATCAT CCTCTGCATG	1200
10	GTCAGGTCAT GGATGAGCAG ACGATGGTGC AGGATATCCT GCTGATGAAG CAGAACAAC	1260
	TTAACGCCGT GCGCTGTTTG CATTATCCGA ACCATCCGCT GTGGTACACG CTGTGCGACC	1320
	GCTACGGCCT GTATGTGGTG GATGAAGCCA ATATTGAAAC CCACGGCATG GTGCCAATGA	1380
	ATCGTCTGAC CGATGATCCG CGCTGGCTAC CGGCGATGAG CGAACGCGTA ACGCGAATGG	1440
	TGCAGCGCGA TCGTAATCAC CCGAGTGTGA TCATCTGGTC GCTGGGGAAT GAATCAGGCC	1500
15	ACGGCGCTAA TCACGACGCG CTGTATCGCT GGATCAAATC TGTCGATCCT TCCCGCCCGG	1560
	TGCAGTATGA AGGCGGCGGA GCCGACACCA CGGCCACCGA TATTATTTGC CCGATGTACG	1620
	CGCGCGTGGA TGAAGACCAG CCCTTCCCGG CTGTGCCGAA ATGGTCCATC AAAAAATGGC	1680
	TTTCGCTACC TGGAGAGACG CGCCCGCTGA TCCTTTGCGA ATACGCCAC GCGATGGGTA	1740
	ACAGTCTTGG CGGTTTCGCT AAATACTGGC AGGCGTTTCG TCAGTATCCC CGTTTACAGG	1800
20	GCGGCTTCGT CTGGGACTGG GTGGATCAGT CGCTGATTAA ATATGATGAA AACGGCAACC	1860
	CGTGCTCGGC TTACGGCGGT GATTTTGGCG ATACGCCGAA CGATCGCCAG TTCTGTATGA	1920
	ACGGTCTGGT CTTTGCCGAC CGCACGCCGC ATCCAGCGCT GACGGAAGCA AAACACCAGC	1980
	AGCAGTTTTT CCAGTTCCGT TTATCCGGG AAACCATCGA AGTGACCAGC GAATACCTGT	2040
	TCCGTCATAG CGATAACGAG CTCCTGCACT GGATGGTGGC GCTGGATGGT AAGCCGCTGG	2100
25	CAAGCGGTGA AGTGCCTCTG GATGTCGCTC CACAAGGTAA ACAGTTGATT GAACTGCCTG	2160
	AACTACCGCA GCCGGAGAGC GCCGGGCAAC TCTGGCTCAC AGTACGCGTA GTGCAACCGA	2220
	ACGCGACCGC ATGGTCAGAA GCCGGGCACA TCAGCGCCTG GCAGCAGTGG CGTCTGGCGG	2280
	AAAACCTCAG TGTGACGCTC CCCGCCCGCT CCCACGCCAT CCCGCATCTG ACCACCAGCG	2340
	AAATGGATTT TTGCATCGAG CTGGGTAATA AGCGTTGGCA ATTTAACCGC CAGTCAGGCT	2400
30	TTCTTTTACA GATGTGGATT GGCGATAAAA AACAACTGCT GACGCCGCTG CGCGATCAGT	2460
	TCACCCGTGC ACCGCTGGAT AACGACATTG GCGTAAGTGA AGCGACCCGC ATTGACCCTA	2520
	ACGCCTGGGT CGAACGCTGG AAGGCGGCGG GCCATTACCA GGCCGAAGCA GCGTTGTTGC	2580
	AGTGACCGC AGATACACTT GCTGATGCGG TGCTGATTAC GACCGCTCAC GCGTGGCAGC	2640
	ATCAGGGGAA AACCTTATTT ATCAGCCGGA AAACCTACCG GATTGATGGT AGTGGTCAAA	2700
35	TGGCGATTAC CGTTGATGTT GAAGTGGCGA GCGATACACC GCATCCGGCG CGGATTGGCC	2760
	TGAACTGCCA GCTGGCGCAG GTAGCAGAGC GGGTAAACTG GCTCGGATTA GGGCCGCAAG	2820
	AAAACATATC CGACCGCCTT ACTGCCGCCT GTTTTGACCG CTGGGATCTG CCATTGTCAG	2880
	ACATGTATAC CCCGTACGTC TTCCCGAGCG AAAACGGTCT GCGCTGCGGG ACGCGCGAAT	2940

5	TGAATTATGG CCCACACCAG TGGCGCGGCG ACTTCCAGTT CAACATCAGC CGCTACAGTC	3000
	AACAGCAACT GATGGAAACC AGCCATCGCC ATCTGCTGCA CGCGGAAGAA GGCACATGGC	3060
	TGAATATCGA CGGTTTCCAT ATGGGGATTG GTGGCGACGA CTCCTGGAGC CCGTCAGTAT	3120
	CGGCGGATTC CAGCTGAGCG CCGGTCGCTA CCATTACCAG TTGGTCTGGT GTCAAAAATA	3180
	ATAATAACCG GGCAGGCCAT GTCTGCCCCG ATTTTCGCGTA AGGAAATCCA TTATGTACTA	3240
10	TTTAAAAAAC ACAAACTTTT GGATGTTCCG TTTATTCTTT TTCTTTTACT TTTTTATCAT	3300
	GGGAGCCTAC TTCCCGTTTT TCCCGATTTC GCTACATGAC ATCAACCATA TCAGCAAAAG	3360
	TGATACGGGT ATTATTTTTG CCGCTATTTT TCTGTTCTCG CTATTATTCC AACCGCTGTT	3420
	TGGTCTGCTT TCTGACAAAC TCGGGCTGCG CAAATACCTG CTGTGGATTA TTACCGGCAT	3480
	GTTAGTGATG TTTGCGCCGT TCTTTATTTT TATCTTCGGG CCACTGTTAC AATACAACAT	3540
15	TTTAGTAGGA TCGATTGTTG GTGGTATTTA TCTAGGCTTT TGTTTTAACG CCGGTGCGCC	3600
	AGCAGTAGAG GCATTTATTG AGAAAGTCAG CCGTCGCAGT AATTTCGAAT TTGGTTCGCGC	3660
	GCGGATGTTT GGCTGTGTTG GCTGGGCGCT GTGTGCCTCG ATTGTTCGGCA TCATGTTTAC	3720
	CATCAATAAT CAGTTTGTTT TCTGGCTGGG CTCTGGCTGT GCACTCATCC TCGCCGTTTT	3780
	ACTCTTTTTT GCCAAAACGG ATGCGCCCTC TTCTGCCACG GTTGCCAATG CGGTAGGTGC	3840
20	CAACCATTTC GCATTTAGCC TTAAGCTGGC ACTGGAACG TTCAGACAGC CAAAACGTG	3900
	GTTTTTGTC CTGTATGTTA TTGGCGTTTC CTGCACCTAC GATGTTTTTG ACCAACAGTT	3960
	TGCTAATTTT TTTACTTCGT TCTTTGCTAC CCGTGAACAG GGTACGCGGG TATTTGGCTA	4020
	CGTAACGACA ATGGGCGAAT TACTTAACGC CTCGATTATG TTCTTTGCGC CACTGATCAT	4080
	TAATCGCATC GGTGGGAAAA ACGCCCTGCT GCTGGCTGGC ACTATTATGT CTGTACGTAT	4140
25	TATTGGCTCA TCGTTCGCCA CCTCAGCGCT GGAAGTGGTT ATTCTGAAAA CGCTGCATAT	4200
	GTTTGAAAGTA CCGTTCCTGC TGGTGGGCTG CTTTAAATAT ATTACCAGCC AGTTTGAAGT	4260
	GCGTTTTTCA GCGACGATTT ATCTGGTCTG TTTCTGCTTC TTTAAGCAAC TGGCGATGAT	4320
	TTTTATGTCT GTACTGGCGG GCAATATGTA TGAAAGCATC GGTTCACAGG GCGCTTATCT	4380
	GGTGCTGGGT CTGGTGGCGC TGGGCTTCAC CTTAATTTCC GTGTTCACGC TTAGCGGCCC	4440
30	CGGCCCCGTT TCCCTGCTGC GTCGTCAGGT GAATGAAGTC GCTTAAGCAA TCAATGTCGG	4500
	ATGCGGCGCG ACGCTTATCC GACCAACATA TCATAACGGA GTGATCGCAT TGAACATGCC	4560
	AATGACCGAA AGAATAAGAG CAGGCAAGCT ATTTACCGAT ATGTGCGAAG GCTTACCGGA	4620
	AAAAAGACTT CGTGGGAAAA CGTTAATGTA TGAGTTTAAT CACTCGCATC CATCAGAAGT	4680
	TGAAAAAGA GAAAGCCTGA TTAAAGAAAT GTTTGCCACG GTAGGGGAAA ACGCCTGGGT	4740
35	AGAACCGCCT GTCTATTTCT CTTACGGTTC CAACATCCAT ATAGGCCGCA ATTTTTATGC	4800
	AAATTTCAAT TTAACCATTG TCGATGACTA CACGGTAACA ATCGGTGATA ACGTACTGAT	4860
	TGCACCCAAC GTTACTCTTT CCGTTACGGG ACACCCTGTA CACCATGAAT TGAGAAAAA	4920
	CGGCGAGATG TACTCTTTTC CGATAACGAT TGGCAATAAC GTCTGGATCG GAAGTCATGT	4980

5	GGTTATTAAT	CCAGGCGTCA	CCATCGGGGA	TAATTCTGTT	ATTGGCGCGG	GTAGTATCGT	5040
	CACAAAAGAC	ATTCCACCAA	ACGTCGTGGC	GGCTGGCGTT	CCTTGTCGGG	TTATTTCGCGA	5100
	AATAAACGAC	CGGGATAAGC	ACTATTATTT	CAAAGATTAT	AAAGTTGAAT	CGTCAGTTTA	5160
	AATTATAAAA	ATTGCCTGAT	ACGCTGCGCT	TATCAGGCCT	ACAAGTTCAG	CGATCTACAT	5220
	TAGCCGCATC	CGGCATGAAC	AAAGCGCAGG	AACAAGCGTC	GCATCATGCC	TCTTTGACCC	5280
10	ACAGCTGCGG	AAAACGTACT	GGTGCAAAAC	GCAGGGTTAT	GATCATCAGC	CCAACGACGC	5340
	ACAGCGCATG	AAATGCCCAG	TCCATCAGGT	AATTGCCGCT	GATACTACGC	AGCACGCCAG	5400
	AAAACCACGG	GGCAAGCCCC	GCGATGATAA	AACCGATTCC	CTGCATAAAC	GCCACCAGCT	5460
	TGCCAGCAAT	AGCCGGTTGC	ACAGAGTGAT	CGAGCGCCAG	CAGCAAACAG	AGCGGAAACG	5520
	CGCCGCCCAG	ACCTAACCCA	CACACCATCG	CCCACAATAC	CGGCAATTGC	ATCGGCAGCC	5580
15	AGATAAAGCC	GCAGAACCCC	ACCAGTTGTA	ACACCAGCGC	CAGCATTAAC	AGTTTGCGCC	5640
	GATCCTGATG	GCGAGCCATA	GCAGGCATCA	GCAAAGCTCC	TGCGGCTTGC	CCAAGCGTCA	5700
	TCAATGCCAG	TAAGGAACCG	CTGTACTGCG	CGCTGGCACC	AATCTCAATA	TAGAAAGCGG	5760
	GTAACCAGGC	AATCAGGCTG	GCGTAACCGC	CGTTAATCAG	ACCGAAGTAA	ACACCAGCG	5820
	TCCACGCGCG	GGGAGTGAAT	ACCACGCGAA	CCGGAGTGGT	TGTTGTCTTG	TGGGAAGAGG	5880
20	CGACCTCGCG	GGCGCTTTGC	CACCACCAGG	CAAAGAGCGC	AACAACGGCA	GGCAGCGCCA	5940
	CCAGGCGAGT	GTTTGATACC	AGGTTTCGCT	ATGTTGAACT	AACCAGGGCG	TTATGGCGGC	6000
	ACCAAGCCCA	CCGCCGCCCA	TCAGAGCCGC	GGACCACAGC	CCCATCACCA	GTGGCGTGCG	6060
	CTGCTGAAAC	CGCCGTTTAA	TCACCGAAGC	ATCACCGCCT	GAATGATGCC	GATCCCCACC	6120
	CCACCAAGCA	GTGCGCTGCT	AAGCAGCAGC	GCACTTTGCG	GGTAAAGCTC	ACGCATCAAT	6180
25	GCACCGACGG	CAATCAGCAA	CAGACTGATG	GCGACACTGC	GACGTTTCGCT	GACATGCTGA	6240
	TGAAGCCAGC	TTCCGGCCAG	CGCCAGCCCG	CCCATGGTAA	CCACCGGCAG	AGCGGTCGAC	6300
	CCGGACGGGA	CGCTCCTGCG	CCTGATACAG	AACGAATTGC	TTGCAGGCAT	CTCATGAGTG	6360
	TGTCTTCCCG	TTTTCCGCCT	GAGGTCACTG	CGTGGATGGA	GCGCTGGCGC	CTGCTGCGCG	6420
	ACGGCGAGCT	GCTCACCACC	CACTCGAGCT	GGATACTTCC	CGTCCGCCAG	GGGGACATGC	6480
30	CGGCGATGCT	GAAGGTCGCG	CGCATTCCCC	ATGAAGAGGC	CGGTTACCGC	CTGTTGACCT	6540
	GGTGGGACGG	GCAGGGCGCC	GCCCCAGTCT	TCGCCTCGGC	GGCGGGCGCT	CTGCTCATGG	6600
	AGCGCGCGTC	CGGGGCCCGG	GACCTTGAC	AGATAGCGTG	GTCCGGCCAG	GACGACGAGG	6660
	CTTGCAGGAT	CTATGATTCC	CTTTGTCAAC	AGCAATGGAT	CACTGAAAAT	GGTTCAATGA	6720
	TCACATTAAG	TGGTATTCAA	TATTTTCATG	AAATGGGAAT	TGACGTTCCCT	TCCAAACATT	6780
35	CACGTAAAAT	CTGTTGTGCG	TGTTTAGATT	GGAGTGAACG	CCGTTTCCAT	TTAGGTGGGT	6840
	ACGTTGGAGC	CGCATTATTT	TCGCTTTATG	AATCTAAAGG	GTGGTTAACT	CGACATCTTG	6900
	GTTACCGTGA	AGTTACCATC	ACGGAAAAAG	GTTATGCTGC	TTTAAAGACC	CACTTTTACA	6960
	TTTAAGTTGT	TTTTCTAATC	CGCATATGAT	CAATTCAAGG	CCGAATAAGA	AGGCTGGCTC	7020

5	TGCACCTTGG	TGATCAAATA	ATTCGATAGC	TTGTCGTAAT	AATGGCGGCA	TACTATCAGT	7080
	AGTAGGTGTT	TCCCTTTCTT	CTTTAGCGAC	TTGATGCTCT	TGATCTTCCA	ATACGCAACC	7140
	TAAAGTAAAA	TGCCCCACAG	CGCTGAGTGC	ATATAATGCA	TTCTCTAGTG	AAAAACCTTG	7200
	TTGGCATAAA	AAGGCTAATT	GATTTTCGAG	AGTTTCATAC	TGTTTTTCTG	TAGGCCGTGT	7260
	ACCTAAATGT	ACTTTTGCTC	CATCGCGATG	ACTTAGTAAA	GCACATCTAA	AACTTTTAGC	7320
10	GTTATTACGT	AAAAAATCTT	GCCAGCTTTC	CCCTTCTAAA	GGGCAAAAGT	GAGTATGGTG	7380
	CCTATCTAAC	ATCTCAATGG	CTAAGGCGTC	GAGCAAAGCC	CGCTTATTTT	TTACATGCCA	7440
	ATACAATGTA	GGCTGCTCTA	CACCTAGCTT	CTGGGCGAGT	TTACGGGTTG	TTAAACCTTC	7500
	GATTCCGACC	TCATTAAGCA	GCTCTAATGC	GCTGTTAATC	ACTTTACTTT	TATCTAATCT	7560
	AGACATCATT	AATTCCTAAT	TTTTGTTGAC	ACTCTATCAT	TGATAGAGTT	ATTTTACCAC	7620
15	TCCCTATCAG	TGATAGAGAA	AAGTGAAATG	AATAGTTCGA	CAAAGATCGC	ATTGGTAATT	7680
	ACGTTACTCG	ATGCCATGGG	GATTGGCCTT	ATCATGCCAG	TCTTGCCAAC	GTTATTACGT	7740
	GAATTTATTG	CTTCGGAAGA	TATCGCTAAC	CACTTTGGCG	TATTGCTTGC	ACTTTATGCG	7800
	TTAATGCAGG	TTATCTTTGC	TCCTTGCTT	GGAAAAATGT	CTGACCGATT	TGGTCGGCGC	7860
	CCAGTGCTGT	TGTTGTCATT	AATAGGCGCA	TCGCTGGATT	ACTTATTGCT	GGCTTTTTCA	7920
20	AGTGCCTTT	GGATGCTGTA	TTTAGGCCGT	TTGCTTTCAG	GGATCACAGG	AGCTACTGGG	7980
	GCTGTCGCGG	CATCGGTCAT	TGCCGATACC	ACCTCAGCTT	CTCAACGCGT	GAAGTGGTTC	8040
	GGTTGGTTAG	GGGCAAGTTT	TGGGCTTGGT	TTAATAGCGG	GGCCTATTAT	TGGTGGTTTT	8100
	GCAGGAGAGA	TTTCACCGCA	TAGTCCCTTT	TTTATCGCTG	CGTTGCTAAA	TATTGTCACT	8160
	TTCCTTGTGG	TTATGTTTTG	GTTCCGTGAA	ACCAAAAATA	CACGTGATAA	TACAGATACC	8220
25	GAAGTAGGGG	TTGAGACGCA	ATCGAATTCG	GTATACATCA	CTTTATTTAA	AACGATGCCC	8280
	ATTTTGTTGA	TTATTTATTT	TTCAGCGCAA	TTGATAGGCC	AAATTCCCGC	AACGGTGTGG	8340
	GTGCTATTTA	CCGAAAATCG	TTTTGGATGG	AATAGCATGA	TGGTTGGCTT	TTCATTAGCG	8400
	GGTCTTGGTC	TTTTTACTCT	AGTATTCCAA	GCCTTTGTGG	CAGGAAGAAT	AGCCACTAAA	8460
	TGGGGCGAAA	AAACGGCAGT	ACTGCTCGAA	TTTATTGCAG	ATAGTAGTGC	ATTTGCCTTT	8520
30	TTAGCGTTTA	TATCTGAAGG	TTGGTTAGAT	TTCCCTGTTT	TAATTTTATT	GGCTGGTGGT	8580
	GGGATCGCTT	TACCTGCATT	ACAGGGAGTG	ATGTCTATCC	AAACAAAGAG	TCATGAGCAA	8640
	GGTGCTTTAC	AGGGATTATT	GGTGAGCCTT	ACCAATGCAA	CCGGTGTATT	TGGCCCATTA	8700
	CTGTTTACTG	TTATTTATAA	TCATTCACTA	CCAATTTGGG	ATGGCTGGAT	TTGGATTATT	8760
	GGTTTAGCGT	TTTACTGTAT	TATTATCCTG	CTATCGATGA	CCTTCATGTT	AACCCCTCAA	8820
35	GCTCAGGGGA	GTAAACAGGA	GACAAGTGCT	TAGTTATTTT	GTCACCAAAT	GATGTTATTC	8880
	CGCGAAATAT	AATGACCCTC	TTGATAACCC	AAGAGGGCAT	TTTTTACGAT	AAAGAAGATT	8940
	TAGCTTCAAA	TAAAACCTAT	CTATTTTATT	TATCTTTCAA	GCTCAATAAA	AAGCCGCGGT	9000
	AAATAGCAAT	AAATTGGCCT	TTTTTATCGG	CAAGCTCTTT	TAGGTTTTTC	GCATGTATTG	9060

5	CGATATGCAT	AAACCAGCCA	TTGAGTAAGT	TTTTAAGCAC	ATCACTATCA	TAAGCTTTAA	9120
	GTTGGTTCTC	TTGGATCAAT	TTGCTGACAA	TGGCGTTTAC	CTTACCAGTA	ATGTATTCAA	9180
	GGCTAATTTT	TTCAAGTTCA	TTCCAACCAA	TGATAGGCAT	CACCTTCTTG	ATAGGGATAA	9240
	GGTTTTTATT	ATTATCAATA	ATATAATCAA	GATAATGTTC	AAATATACTT	TCTAAGGCAG	9300
	ACCAACCATT	TGTAAATCA	GTTTTTGTG	TGATGTAGGC	ATCAATCATA	ATTAATTGCT	9360
10	GCTTATAACA	GGCACTGAGT	AATTGTTTTT	TATTTTTTAA	GTGATGATAA	AAGGCACCTT	9420
	TGGTCACCAA	CGCTTTTCCC	GAGATCCTCT	GCGACACCGC	CGCTCGTCTG	CACGCGCCGC	9480
	GGTCCGGACC	GCCGCCCGAT	CTCCATCCGC	TACAGGAATG	GTTCCAGCCG	CTTTTCCGGT	9540
	TGGCCGCTGA	GCACGCGGCA	CTTGCGCCCG	CCGCCAGCGT	AGCGCGCCAA	CTTCTGGCGG	9600
	CGCCGCGCGA	GGTGTGCCCC	CTCCACGGCG	ACCTGCACCA	CGAGAACGTG	CTCGACTTCG	9660
15	GCGACCGCGG	CTGGCTGGCC	ATCGACCCGC	ACGGACTGCT	CGGCGAGCGC	ACCTTCGACT	9720
	ATGCCAACAT	CTTCACGAAT	CCCGATCTCA	GCGACCCCGG	TCGCCCCGCTT	GCGATCCTGC	9780
	CGGGCAGGCT	GGAGGCTCGA	CTCAGCATTG	TGGTCGCGAC	GACCGGGTTT	GAGCCCGAAC	9840
	GGCTTCTTCG	CTGGATCATT	GCATGGACGG	GCTTGTGCGC	AGCCTGGTTC	ATCGGCGACG	9900
	GCGACGGCGA	GGGCGAGGGC	GCTGCGATTG	ATCTGGCCGT	AAACGCCATG	GCACGCCGGT	9960
20	TGCTTGACTA	GCGCGGTCAC	CGATCTCACC	TGGTCGTCGA	GCTAGGTCAG	GCCGTGTCGG	10020
	GCGTGATCCG	CTGGAAGTCG	TTGCGGGCCA	CACCCGCCGC	CTCGAAGCCC	TGCACCAGGC	10080
	CGGCATCGTG	GTGTGCGTGG	CCGAGGGACT	ATGGAAGGTG	CCGGACGATC	TGCCCAGCA	10140
	GGGCCGCCGC	TATGACGCCC	AGCGTCTTGG	TGGCGTGACG	GTGGAGCTGA	AATCGCACCT	10200
	GCCCATCGAG	CGGCAGGCCC	GCGTGATCGG	TGCCACCTGG	CTTGACCAGC	AGTTGATCGA	10260
25	CGGTGGCTCG	GGCTTGGGCG	ACCTGGGCTT	TAGCAGTGAG	GCCAAGTAGG	CGATACAGCA	10320
	GCGCGCGGAC	TTCCTGGCCG	AACAGGGACT	GGCCGAGCGG	CGCGGGCAGC	GCGTGATCCT	10380
	CACCGGAATC	TGCTGGGCAG	CAGCGGGCTC	GGGAACTGGC	GCAGGCCGCG	AAGGACATTG	10440
	CCGCCGATAC	CGGCCTGGAG	CATCGCCCCG	TGGCCGACGG	CCAGCGCGTT	GCCGGCGTCT	10500
	ACCGGCGCCC	CGTCATGCTC	GCCAGCGGGC	GAAATGGGAT	GCTTGATGAC	GCCAAGGGGT	10560
30	CCAGCCTCGT	GCGGTGGAAG	CCCATCGAAC	AGCGGCTTGG	GGAGCAGCTC	GCCGCGACGG	10620
	TGCGCGGTGG	CGGCGTGTCT	TGGGAGATTG	GACGACAGCG	TGGGCCGGCC	CCTGTCTCTT	10680
	GATCAGATCT	TGATCCCCTG	CGCCATCAGA	TCCTTGGCGG	CAAGAAAGCC	ATCCAGTTTA	10740
	CTTTGCAGGG	CTTCCCAACC	TTCCAGAGG	GCGCCCCAGC	TGGCAATTCC	GGTTCGCTTG	10800
	CTGTCCATAA	AACCGCCAG	TCTAGCTATC	GCCATGTAAG	CCCACTGCAA	GCTACCTGCT	10860
35	TTCTCTTTGC	GCTTGCGTTT	TCCCTTGTCC	AGATAGCCCA	GTAGCTGACA	TTCATCCGGG	10920
	GTCAGCACCG	TTTCTGCGGA	CTGGCTTTCT	ACGTGTTCCG	CTTCCTTTAG	CAGCCCTTGC	10980
	GCCCTGAGTG	CTTGCGGCAG	CGTGAAGCTT	TCTCTGAGCT	GTAACAGCCT	GACCGCAACA	11040
	AACGAGAGGA	TCGAGACCAT	CCGCTCCAGA	TTATCCGGCT	CCTCCATGCG	TTGCCTCTCG	11100

5	GCTCCTGCTC	CGGTTTTCCA	TGCCTTATGG	AACTCCTCGA	TCCGCCAGCG	ATGGGTATAA	11160
	ATGTCGATGA	CGCGCAAGGC	TTGGGCTAGC	GACTCGACCG	GTTCCGCCGT	CAGCAACAAC	11220
	CATTTCAACG	GGGTCTCACC	CTTGGGCGGG	TTAATCTCCT	CGGCCAGCAC	CGCGTTGAGC	11280
	GTGATATTCC	CCTGTTTTAG	CGTGATGCGC	CCACTGCGCA	GGCTCAAGCT	CGCCTTGCGG	11340
	GCTGGTCGAT	TTTTACGTTT	ACCGCGTTTA	TCCACCACGC	CCTTTTGCGG	AATGCTGATC	11400
10	TGATAGCCAC	CCAACCTCCG	TTGGTTCTTC	AGATGGTCGA	TCAGATACAA	CCCAGACTCT	11460
	ACGTCCTTGC	GTGGGTGCTT	GGAGCGCACC	ACGAAGCGCT	CGTTATGCGC	CAGCCTGTCC	11520
	TGCAGATAAG	CATGAATATC	GGCTTCGCGG	TCACAGACCG	CAATCACGTT	GCTCATCATG	11580
	CTGCCCATGC	GTAACCGGCT	AGTTGCGGCC	GCTGCCAGCC	ATTTGCCACT	CTCCTTTTCA	11640
	TCCGCATCGG	CAGGGTCATC	CGGGCGCATC	CACCACTCCT	GATGCAGTAA	TCCTACGGTG	11700
15	CGGAATGTGG	TGGCCTCGAG	CAAGAGAACG	GAGTGAACCC	ACCATCCGCG	GGATTTATCC	11760
	TGAATAGAGC	CCAGCTTGCC	AAGCTCTTCG	GCGACCTGGT	GGCGATAACT	CAAAGAGGTG	11820
	GTGTCCTCAA	TGGCCAGCAG	TTCGGGAAAC	TCCTGAGCCA	ACTTGACTGT	TTGCATGGCG	11880
	CCAGCCTTTC	TGATCGCCTC	GGCAGAAACG	TTGGGATTGC	GGATAAATCG	GTAAGCGCCT	11940
	TCCTGCATGG	CTTCACTACC	CTCTGATGAG	ATGGTTATTG	ATTTACCAGA	ATATTTTGCC	12000
20	AATTGGGCGG	CGACGTTAAC	CAAGCGGGCA	GTACGGCGAG	GATCACCCAG	CGCCGCCGAA	12060
	GAGAACACAG	ATTTAGCCCA	GTCGGCCGCA	CGATGAAGAG	CAGAAGTTAT	CATGAACGTT	12120
	ACCATGTTAG	GAGGTCACAT	GGAAGATCAG	ATCCTGGAAA	ACGGGAAAGG	TTCCGTTCTGA	12180
	ATTGCATGCG	GATCCGGGAT	CAAGATCTGA	TCAAGAGACA	GGTACCAATT	GTTGAAGACG	12240
	AAAGGGCCTC	GTGATACGCC	TATTTTTATA	GGTTAATGTC	ATGATAATAA	TGGTTTCTTA	12300
25	GACGTCAGGT	GGCACTTTTC	GGGGAAATGT	GCGCGGAACC	CCTATTTGTT	TATTTTTCTA	12360
	AATACATTCA	AATATGTATC	CGCTCATGAG	ACAATAACCC	TGATAAATGC	TTCAATAATA	12420
	TTGAAAAAGG	AAGAGTATGA	GTATTCAACA	TTTCCGTGTC	GCCCTTATTC	CCTTTTTTGC	12480
	GGCATTTTGC	CTTCCTGTTT	TTGCTCACCC	AGAAACGCTG	GTGAAAGTAA	AAGATGCTGA	12540
	AGATCAGTTG	GGTGCACGAG	TGGGTTACAT	CGAACTGGAT	CTCAACAGCG	GTAAGATCCT	12600
30	TGAGAGTTTT	CGCCCCGAAG	AACGTTTTCC	AATGATGAGC	ACTTTTAAAG	TTCTGCTATG	12660
	TGGCGCGGTA	TTATCCCGTG	TTGACGCCGG	GCAAGAGCAA	CTCGGTCGCC	GCATACACTA	12720
	TTCTCAGAAT	GACTTGTTTG	AGTACTTGGC	AAACTGATCT	AAATGTTTAG	CCCAGTCATC	12780
	ATACTTCACC	GATGCCAACG	CATTAAAAAT	AGCATCACGA	TCGGCTTTGC	TGAATTTCTT	12840
	ATTTAAAAACA	TCCTTGTTAT	TTTCAAAAGC	AGCGAGAGCT	TCATTCACAT	TGCCGATTTT	12900
35	CTTACCTTTA	GACTTATCAG	CAAGTTCCTG	TGCCATTTTC	GAATATTTTT	CACCATATTT	12960
	TTCAGTCAGC	GTTTGATAAA	AGCTAACTGT	TGCATCAACA	GCATCCTTAA	TCTGTGAATT	13020
	AAGGAGATTA	TTCTGTGCTT	TTTTCAAATT	TTCTTCAGCT	TCATGAACAC	GAGCGATACC	13080
	GGCATTACGA	TTATTACTGA	CCTGAGAAAT	AGCCTTCTGG	ATCTGAGTTA	TATCAGCATT	13140

5	TATCCGGTTA	ATACGTGTTT	CTGATGCTGT	TACCTGTTTT	TGTTTTTCTT	CTCTAATCTT	13200
	ACCGGCCCCA	ACCCGTCGTC	TGGTTGCTTC	AAAAAAGGA	CGGTTCTGAA	GCGGATCATT	13260
	GGCTCTTGGT	GATAGTTTTT	TGACCAGCTC	ATCCAGTTCT	TTATATTTAG	CGGATGCCTG	13320
	AGCCAGTTCA	TTTCGTTTTT	CAGCGAGCGT	TTTCATTTCT	GCATCACGGG	CATGGATACT	13380
	GGAGCTTAAA	CGAGAATTGA	GAGTCTTAAT	CTCTCCATCC	ATTTTCACCA	CTTCAGATTG	13440
10	TGCAGCAGAA	AGTTTTTTTT	GGGCGATCTC	AACAGCTTTA	GCTTCTTCAC	TCAATGCAGC	13500
	CAGTCGTTTC	TCTTCAGCTT	CAGCCAGTTT	CAACTGGCGT	TCTGTTTCAG	CCTTCTCCCG	13560
	TTCAATCTCT	TTACGTCGTT	GTTCTGCTTC	CTGAAAAGCC	TTTTCTGCTG	CTTCCGCTTC	13620
	TTTACGGGCT	TTTTCTTCTG	CTTTCGCAAG	GCGCAAACGC	TCTGCTTCCG	CCTGCATAGC	13680
	TGCATTATTA	GCATGAGCAA	GCTCTGTTGC	TGAAGGCGTA	CGTGAGGCAT	TGTGACGAAG	13740
15	AGCCTCATTC	ACGATATCCT	TCAGGCGCTG	AGTCAGCGCA	TCCCTGTTTG	CCTTTGCTTT	13800
	CGCCTGTGCT	TCCGCTGCAG	CTTTTGCCCG	GGCAGCCTGC	TCTGCCTGTG	TTTTCTTTAA	13860
	TTGAGCAGTA	GACCATTTAG	CAGTTGCATG	AATAGCTGCA	GAACTTTCAC	TTTTACTGCC	13920
	TCCTTTTCCA	CCTCCGCCGC	CAGAGCCACT	CCCGTCAGGA	GTACCATTCA	AAAGAGTAAT	13980
	AATTACCTGT	CCCTTATCAT	CATAAGGAAC	ACCATCTTTA	TAGTACGCTA	CCGCGGTTTC	14040
20	CATTATAAAA	TCCTCTTTGA	CTTTTAAAC	AATAAGTTAA	AAATAAATAC	TGTACATATA	14100
	ACCACTGGTT	TTATATACAG	CATAAAAGCT	ACGCCGCTGC	ATTTTCCCTG	TCAAGACTGT	14160
	GGACTTCCAT	TTTTGTGAAA	ACGATCAAAA	AAACAGTCTT	TCACACCACG	CGCTATTCTC	14220
	GCCCGATGCC	ACAAAAACCA	GCACAAACAT	TACCGTTCTC	AGACCTCATT	ATGTTTTACT	14280
	GAAACTATGA	GATGAGACAT	CTATGGGACA	CTGTCACTTT	ATGGCATGGC	ACACACTCCG	14340
25	GGACGCACTA	AAAATGACAG	GCAGATCGCG	TTACAGTTT	TACCGTGATA	TGCGCGGAGG	14400
	CCTTGTCAGT	TACCGTACCG	GCAGGGACGG	ACGACGGGAG	TTTGAAACCA	GTGAACTGAT	14460
	CCGGGCATAC	GGCGAATTAA	AGCAGAATGA	GACACCAGAA	AGGCACAGTG	AGGGACATGC	14520
	AGAAAATCCA	CATGATCAGC	AGACAGAACG	CATTCTCCGG	GAAGTGAATG	AGCTGAAACA	14580
	ATGCCTGACG	CTGATGCTTG	AGGATAAACA	GGCACAGGAT	ATGGATCGCA	GACGCCAGGA	14640
30	AGCAGAACGG	GAACAGCTAC	AAAATGAGAT	AGCCCAGCTC	AGGCAGGCAC	TGGAAGTGGG	14700
	AAAGAAACGG	GGATTCTGGT	CCAGGTTGTT	CGGTCGCTGA	ACGCTGTCAG	AGACTGATGA	14760
	TAAAATAGTC	TTCGGATAAT	AACTCACCAG	GAATAAATAC	TTTAAGGTAG	GGAGACACTC	14820
	ATGAGACGTA	CCGGAAACAA	ACTTTGTCTT	ATCGCCATGA	TAACAGCAAC	AGTAGCTCTC	14880
	ACAGCCTGTA	CCCCAAAGGG	CAGCGTGGAA	CAACATACCC	GGCATTACGT	ATATGCTTCT	14940
35	GATGACGGTT	TTGATCCCAA	CTTTTCCACC	CAAAAAGCCG	ACACAACACG	AATGATGGTG	15000
	CCTTTTTTTC	GGCAGTTCTG	GGATATGGGA	GCTAAAGACA	AAGCGACAGG	AAAATCACGG	15060
	AGTGATGTGC	AACAACGCAT	TCAGCAGTTT	CACAGCCAAG	AATTTTTTAA	CTCACTCCGG	15120
	GGCACAACTC	AATTTGCGGG	TACTGATTAC	CGCAGCAAAG	ACCTTACCCC	GAAAAAATCC	15180

5	AGGCTGCTGG	CTGACACGAT	TTCTGCGGTT	TATCTCGATG	GCTACGAGGG	CAGACAGTAA	15240
	GTGGATTTAC	CATAATCCCT	TAATTGTACG	CACCGCTAAA	ACGCGTTCAG	CGCGATCACG	15300
	GCAGCAGACA	GGTAAAAATG	GCAACAAACC	ACCCTAAAAA	CTGCGCGATC	GCGCCTGATA	15360
	AATTTTAACC	GTATGAATAC	CTATGCAACC	AGAGGGTACA	GGCCACATTA	CCCCACTTA	15420
	ATCCACTGAA	GCTGCCATTT	TTCATGGTTT	CACCATCCCA	GCGAAGGGCC	ATCCAGCGTG	15480
10	CGTTCCTGTA	TTTCCGGCTG	ACGCTCCCGT	TCTAGGGATA	ACACATGTTT	GCGCTCCTGT	15540
	ATCAGCCGTT	CCTCTCTTAT	CTCCAGTTCT	CGCTGTATAA	CTGGCTCAAG	CGTTCTGTCT	15600
	GCTCGCTCAA	GTGTTGCACC	TGCTGACTCA	ACTGCATGAC	CCGCTCGTTC	AGCATCGCGT	15660
	TGTCCCGTTG	CGTAAGCGAA	AACATCTTCT	GCAATTCCAC	GAAGGCGCTC	TCCCATTTCG	15720
	TCAGCCGCTG	CATATAGTCC	TGTTGCAGCT	GCTCTAAGGC	GTTTCAGCAA	TGTGTTTCCA	15780
15	GCTCTGTGTC	TCTGTGTGTC	TCCTTCAGAT	GTACCCACTC	TTTCCCCTGA	AAGGGAATCA	15840
	CCTCCGCTGA	TTTCCCGTAC	GGAAGGACAA	GGAATTTTCT	GTTCCCCTCC	TGCACAAACT	15900
	CCACGCCCCA	TGTCTTCGCG	TTCAGTTTCT	GCAATGTCTC	TTCCTGCTTC	CTGATTTCTT	15960
	CCAGGTTTCG	CTGTATCCTC	CCTCCAAGAT	ACCAGAGCGT	CCCGCCACTC	GCGGTAAACA	16020
	GGAGAAAGAC	TATCCCCAGT	AACATCATGC	CCGTATTCCC	TGCCAGCTTT	AACACGTCCC	16080
20	TCCTGTGCTG	CATCATCGCC	TCTTTACACC	CTTCCCGGTG	TTTTTCCAGC	GATTCCTCTG	16140
	TCGAGGCTGT	GAACAGGGCT	ATAGCGTCTC	TGATTTTCGT	CTCGTTTGAT	GTCACAGCCT	16200
	CGCTTACAGA	TTCGCCGAGC	CTCCTGAACT	CGTTGTTTCT	CATTTTCTCT	GTAGATTCGG	16260
	CTCTCTCTTT	CAGCTTTTTT	TCGAACTCCG	CGCCCGTCTG	CAAAAGATTG	CTCATAAAAT	16320
	GCTCCTTTCA	GCCTGATATT	CTTCCCGCCG	TTCGGATCTG	CAATGCTGAT	ACTGCTTCGC	16380
25	GTCACCCTGA	CCACTTCCAG	CCCCGCCTCA	GTGAGCGCCT	GAATCACATC	CTGACGGCCT	16440
	TTTATCTCTC	CGGCATGGTA	AAGTGCATCT	ATACCTCGCG	TGACGCCCTC	AGCAAGCGCC	16500
	TGTTTTCGTT	CAGGCAGGTT	ATCAGGGAGT	GTCAGCGTCC	TGCGGTTCTC	CGGGGCGTTC	16560
	GGGTCATGCA	GCCCCGTAAT	GTGATTTAAC	AGCGTCTGCC	AAGCATCAAT	TCTAGGCCTG	16620
	TCTGCGCGGT	CGTAGTACGG	CTGGAGGCGT	TTTCCGGTCT	GTAGCTCCAT	GTTCGGAATG	16680
30	ACAAAATTCA	GCTCAAGCCG	TCCCTTGTCC	TGGTGCTCCA	CCCACAGGAT	GCTGTACTGA	16740
	TTTTTTTCGA	GACCGGGCAT	CAGTACACGC	TCAAAGCTCG	CCATCACTTT	TTCACGTCCT	16800
	CCCGGCGGCA	GCTCCTTCTC	CGCGAACGAC	AGAACACCGG	ACGTGTATTT	CTTCGCAAAT	16860
	GGCGTGGCAT	CGATGAGTTC	CCGGACTTCT	TCCGGTATAC	CCTGAAGCAC	CGTTGCGCCT	16920
	TCGCGGTTAC	GCTCCCTCCC	CAGCAGGTAA	TCAACCGGAC	CACTGCCACC	ACCTTTTCCC	16980
35	CTGGCATGAA	ATTTAACTAT	CATCCCGCGC	CCCCTGTTCC	CTGACAGCCA	GACGCAGCCG	17040
	GCGCAGCTCA	TCCCCGATGG	CCATCAGTGC	GGCCACCACC	TGAACCCGGT	CACCGGAAGA	17100
	CCACTGCCCC	CTGTTACACT	TACGGGCTGT	CTGATTCAGG	TTATTTCCGA	TGGCGGCCAG	17160
	CTGACGCAGT	AACGGCGGTG	CCAGTGTCGG	CAGTTTTTCC	GAACGGGCAA	CCGGCTCCCC	17220

5	CAGGCAGACC CGCCGCATCC ATACCGCCAG TTGTTTACCC TCACAGCGTT CAAGTAACCG	17280
	GGCATGTTCA TCATCAGTAA CCCGTATTGT GAGCATCCTC TCGCGTTTCA TCGGTATCAT	17340
	TACCCCATGA ACAGAAATCC CCCTTACACG GAGGCATCAG TGAATAACA GGAAAAACC	17400
	GCCCTTAACA TGGCCCGCTT TATCAGAAGC CAGACATTAA CGCTGCTGGA GAAGCTCAAC	17460
	GAAGTGGACG CAGATGAACA GGCCGATATT TGTGAATCGC TTCACGACCA CGCCGATGAG	17520
10	CTTTACCGCA GCTGCCTCGC ACGTTTCGGG GATGACGGTG AAAACCTCTG ACACATGCAG	17580
	CTCCCGGAGA CGGTCACAGC TTGTCTGTGA GCGGATGCCG GGAGCTGACA AGCCCGTCAG	17640
	GGCGCGTCAG CAGGTTTTAG CGGGTGTCTGG GCGCGAGCCC TGACCCAGTC ACGTAGCGAT	17700
	AGCGGAGTGT ATACTGGCTT AACCATGCGG CATCAGTGGC GATTGTATGA AAAGTACGCC	17760
	ATGCCGGGTG TGAAATGCCG CACAGATGCG TAAGGAGAAA ATGCACGTCC AGGCGCTTTT	17820
15	CCGCTTCCTC GCTCACTGAC TCGCTACGCT CGGTCGTTTCG ACTGCGGCGA GCGGTACTGA	17880
	CTCACACAAA AACGGTAACA CAGTTATCCA CAGAATCAGG GGATAAGGCC GGAAAGAACA	17940
	TGTGAGCAAA AGACCAGGAA CAGGAAGAAG GCCACGTAGC AGGCGTTTTT CCATAGGCTC	18000
	CGCCCCCTG ACGAGCATCA CAAAAATAGA CGCTCAAGTC AGAGGTGGCG AAACCCGACA	18060
	GGACTATAAA GCTACCAGGC GTTTCCCCCT GGAAGCTCCC TCGTGCGCTC TCCTGTTCCG	18120
20	ACCCTGCCGC TTACCGGATA CCTGTCCGCC TTTCTCCCTT CGGGAAGCGT GCGGCTTTCT	18180
	CATAGCTCAC GCTGTTGGTA TCTCAGTTCG GTGTAGGTCG TTCGCTCCAA GCTGGGCTGT	18240
	GTGCACGAAC CCCCCGTTCA GCCCGACCGC TGCGCCTTAT CCGGTAATA TCGTCTTGAG	18300
	TCCAACCCGG TAAGGCACGC CTTAACGCCA CTGGCAGCAG CCACTGGTAA CCGGATTAGC	18360
	AGAGCGATGA TGGCACAAAC GGTGCTACAG AGTTCCTTGAA GTAGTGGCCC GACTACGGCT	18420
25	ACACTAGAAG GACAGTATTT GGTATCTGCG CTCTGCTGAA GCCAGTTACC TTCGGAAAAA	18480
	GAGTTGGTAG CTCTTGATCC GGCAACAAA CCACCGTTGG TAGCGGTGGT TTTTTTGT	18540
	GCAAGCAGCA GATTACGCGC AGAAAAAAG GATCTCAAGA AGATCCTTTA ATCTTTTCTA	18600
	CTGAACCGCG ATCCCCGTCA GTTTAGAAGA GGAGGATGGT GCGATGGTCC CTCCCTGAAC	18660
	ATCAGGTATA TAGTTAGCCT GACATCCAAC AAGGAGGTTT ATCGCGAATA TTCCACAAA	18720
30	AAATCTTTTC CTCATAACTC GATCCTTATA AAATGAAAAG AATATATGGC GAGGTTTAAT	18780
	TTATGAGCTT AAGATACTAC ATAAAAAATA TTTTATTTGG CCTGTACTGC AACTTATAT	18840
	ATATATACCT TATAACAAA AACAGCGAAG GGTATTATTT CCTTGTGTCA GATAAGATGC	18900
	TATATGCAAT AGTGATAAGC ACTATTCTAT GTCCATATTC AAAATATGCT ATTGAATACA	18960
	TAGCTTTTAA CTTCATAAAG AAAGATTTTT TCGAAAGAAG AAAAAACCTA AATAACGCCC	19020
35	CCGTAGCAAA ATTAAACCTA TTTATGCTAT ATAATCTACT TTGTTTGGTC CTAGCAATCC	19080
	CATTTGGATT GCTAGGACTT TTTATATCAA TAAAGAATAA TTAAATCCCT AACACCTCAT	19140
	TTATAGTATT AAGTTTATTC TTATCAATAT AGGAGCATAG AA	19182

1. A system for transposing a transposable DNA sequence *in vitro*, the system comprising:

a Tn5 transposase modified relative to a wild type Tn5 transposase, the modified transposase comprising a change relative to the wild type Tn5 transposase that causes the modified transposase to bind to Tn5 outside end repeat sequences with greater avidity than the wild type Tn5 transposase, and a change relative to the wild type Tn5 transposase that causes the modified transposase to be less likely than the wild type transposase to assume an inactive multimeric form;

a donor DNA molecule comprising the transposable DNA sequence, the DNA sequence being flanked at its 5'- and 3'-ends by the Tn5 outside end repeat sequences; and

a target DNA molecule into which the transposable element can transpose.

2. A system as claimed in Claim 1 wherein the change that causes the modified transposase to bind with greater avidity is characterized as a substitution mutation at position 54 of the wild type transposase.

3. A system as claimed in Claim 2 wherein position 54 is a lysine.

4. A system as claimed in Claim 1 wherein the change that causes the modified transposase to be less likely to assume an inactive multimeric form is characterized as a substitution mutation at position 372 of the wild type transposase.

5. A system as claimed in Claim 4 wherein position 372 is a proline.

6. A system as claimed in Claim 1 wherein the modified transposase further comprises a substitution mutation at position 56 of the wild type transposase.

7. A system as claimed in Claim 6 wherein position 56 is an alanine.

8. A system as claimed in Claim 1 wherein the donor DNA molecule is flanked at its 5'- and 3'-ends by an 18 or 19 base pair flanking DNA sequence comprising nucleotide A at position 10, nucleotide T at position 11, and nucleotide A at position 12.

9. The system as claimed in Claim 8 wherein the flanking sequence further comprises a nucleotide at position 4 selected from the group consisting of A or T.

10. The system as claimed in Claim 8 wherein the flanking sequence further comprises a nucleotide at position 15 selected from the group consisting of G or C.

11. The system as claimed in Claim 8 wherein the flanking sequence further comprises a nucleotide at position 17 selected from the group consisting of A or T.

12. The system as claimed in Claim 8 wherein the flanking sequence further comprises a nucleotide at position 18 selected from the group consisting of G or C.

13. The system as claimed in Claim 8 wherein the flanking sequence has the sequence 5'-CTGTCTCTTATACACATCT-3'.

14. The system as claimed in Claim 8 wherein the flanking sequence has the sequence 5'-CTGTCTCTTATACAGATCT-3'.

15. A Tn5 transposase modified relative to a wild type Tn5 transposase, the modified transposase comprising:

a change relative to the wild type Tn5 transposase that causes the modified transposase to bind to Tn5 outside end repeat sequences of a donor DNA with greater avidity than the wild type Tn5 transposase; and

a change relative to the wild type Tn5 transposase that causes the modified transposase to be less likely than the wild type transposase to assume an inactive multimeric form.

16. A modified Tn5 transposase as claimed in Claim 15 wherein the change that causes the modified transposase to bind with greater avidity is characterized as a substitution mutation at position 54 of the wild type transposase.

17. A modified Tn5 transposase as claimed in Claim 16 wherein position 54 is a lysine.

18. A modified Tn5 transposase as claimed in Claim 15 wherein the change that causes the modified transposase to be less likely to assume an inactive multimeric form is characterized as a substitution mutation at position 372 of the wild type transposase.

19. A modified Tn5 transposase as claimed in Claim 18 wherein position 372 is a proline.

20. A modified Tn5 transposase as claimed in Claim 15 further comprising a substitution mutation at position 56 of the wild type transposase.

21. A modified Tn5 transposase as claimed in Claim 20 wherein position 56 is alanine.

22. A genetic construct comprising a nucleotide sequence that can encode a Tn5 transposase that both has greater avidity for Tn5 outside end repeats and is less likely to assume an inactive multimeric form than a wild type Tn5 transposase.

23. A genetic construct as claimed in Claim 22 comprising a nucleotide sequence that encodes a lysine residue at amino acid 54 of the transposase.

24. A genetic construct as claimed in Claim 22 comprising a nucleotide sequence that encodes a proline residue at amino acid 372 of the transposase.

25. A genetic construct as claimed in Claim 22 comprising a nucleotide sequence that encodes a lysine residue at amino acid 54 of the transposase and a proline residue at amino acid 372 of the transposase.

26. A genetic construct as claimed in Claim 22 comprising the nucleotide sequence of SEQ ID NO:1.

27. A genetic construct comprising:
a transposable DNA sequence flanked at its 5' and 3' ends by an 18 or 19 base pair flanking DNA sequence comprising nucleotide A at position 10, nucleotide T at position 11, and nucleotide A at position 12.

28. The construct of Claim 27 further comprising, at position 4 of the flanking sequence, a nucleotide selected from the group consisting of T or A.

29. The construct of Claim 27 further comprising, at position 15 of the flanking sequence, a nucleotide selected from the group consisting of G or C.

30. The construct of Claim 27 further comprising, at position 17 of the flanking sequence, a nucleotide selected from the group consisting of T or A.

31. The construct of Claim 27 further comprising, at position 18 of the flanking sequence, a nucleotide selected from the group consisting of G or C.

32. The construct as claimed in Claim 27 wherein the flanking sequence has the sequence 5'-CTGTCTCTTATACACATCT-3'.

33. The construct as claimed in Claim 27 wherein the flanking sequence has the sequence 5'-CTGTCTCTTATACAGATCT-3'.

34. A method for *in vitro* transposition, the method comprising the steps of:

combining a donor DNA molecule that comprises a transposable DNA sequence of interest, the DNA sequence of interest being flanked at its 5'- and 3'-ends by Tn5 outside end repeat sequences, with a target DNA molecule and a Tn5 transposase modified relative to wild type Tn5 transposase in a suitable reaction buffer at a temperature below a physiological temperature until the modified transposase binds to the outside end repeat sequences; and

raising the temperature to a physiological temperature for a period of time sufficient for the enzyme to catalyze *in vitro* transposition,

wherein the modified transposase comprises a change relative to the wild type Tn5 transposase that causes the modified transposase to bind to the Tn5 outside end repeat sequences with greater avidity than the wild type Tn5 transposase, and a change relative to the wild type Tn5 transposase that causes the modified transposase to be less likely than the wild type transposase to assume an inactive multimeric form.

35. A method as claimed in Claim 34 wherein the change that causes the modified transposase to bind with greater avidity is characterized as a substitution mutation at position 54 of the wild type transposase.

36. A method as claimed in Claim 35 wherein position 54 is a lysine.

37. A method as claimed in Claim 34 wherein the change that causes the modified transposase to be less likely to assume an inactive multimeric form is characterized as a substitution mutation at position 372 of the wild type transposase.

38. A method as claimed in Claim 37 wherein position 372 is a proline.

39. A method as claimed in Claim 34 wherein the modified transposase further comprises a substitution mutation at position 56 of the wild type transposase.

40. A method as claimed in Claim 39 wherein position 56 is an alanine.

41. A method as claimed in Claim 34 wherein the DNA sequence of interest is flanked at its 5'- and 3'-ends by an 18 or 19 base pair flanking DNA sequence comprising nucleotide A at position 10, nucleotide T at position 11, and nucleotide A at position 12.

42. The method as claimed in Claim 41 wherein the flanking sequence further comprises a nucleotide at position 4 selected from the group consisting of A or T.

43. The method as claimed in Claim 41 wherein the flanking sequence further comprises a nucleotide at position 15 selected from the group consisting of G or C.

44. The method as claimed in Claim 41 wherein the flanking sequence further comprises a nucleotide at position 17 selected from the group consisting of A or T.

45. The method as claimed in Claim 41 wherein the flanking sequence further comprises a nucleotide at position 18 selected from the group consisting of G or C.

46. The method as claimed in Claim 41 wherein the flanking sequence has the sequence 5'-CTGTCTCTTATACACATCT-3'.

47. The method as claimed in Claim 41 wherein the flanking sequence has the sequence 5'-CTGTCTCTTATACAGATCT-3'.

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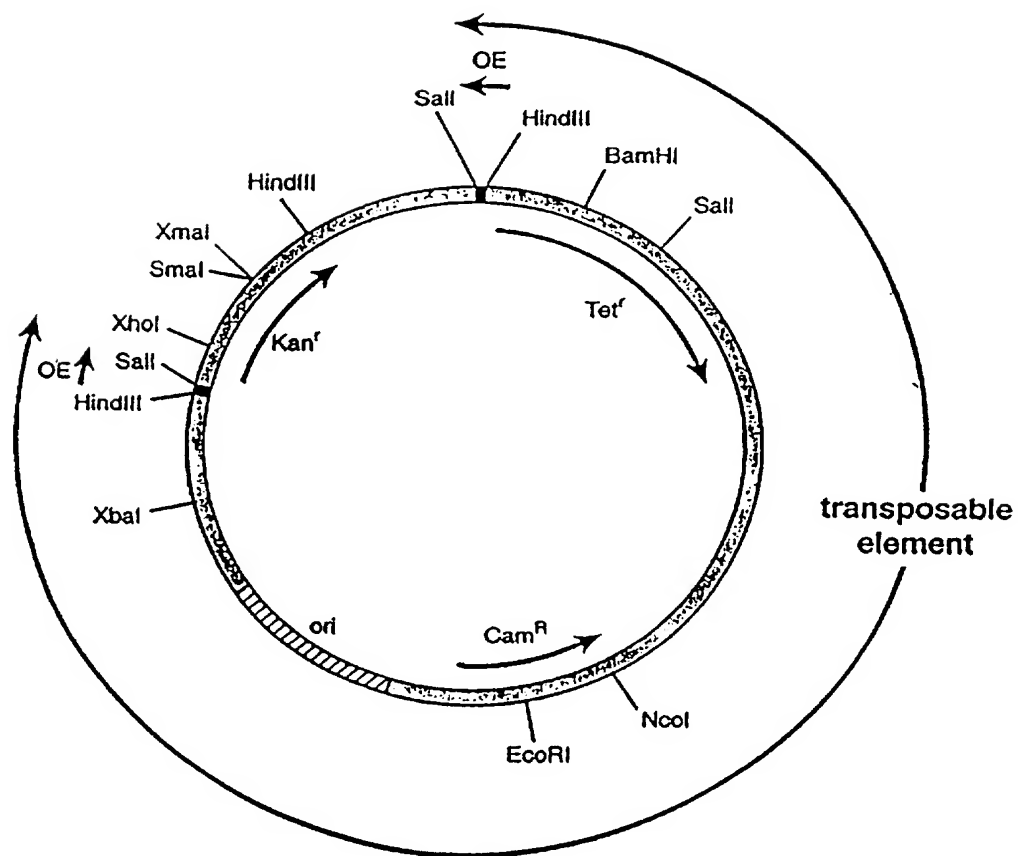


FIG 1

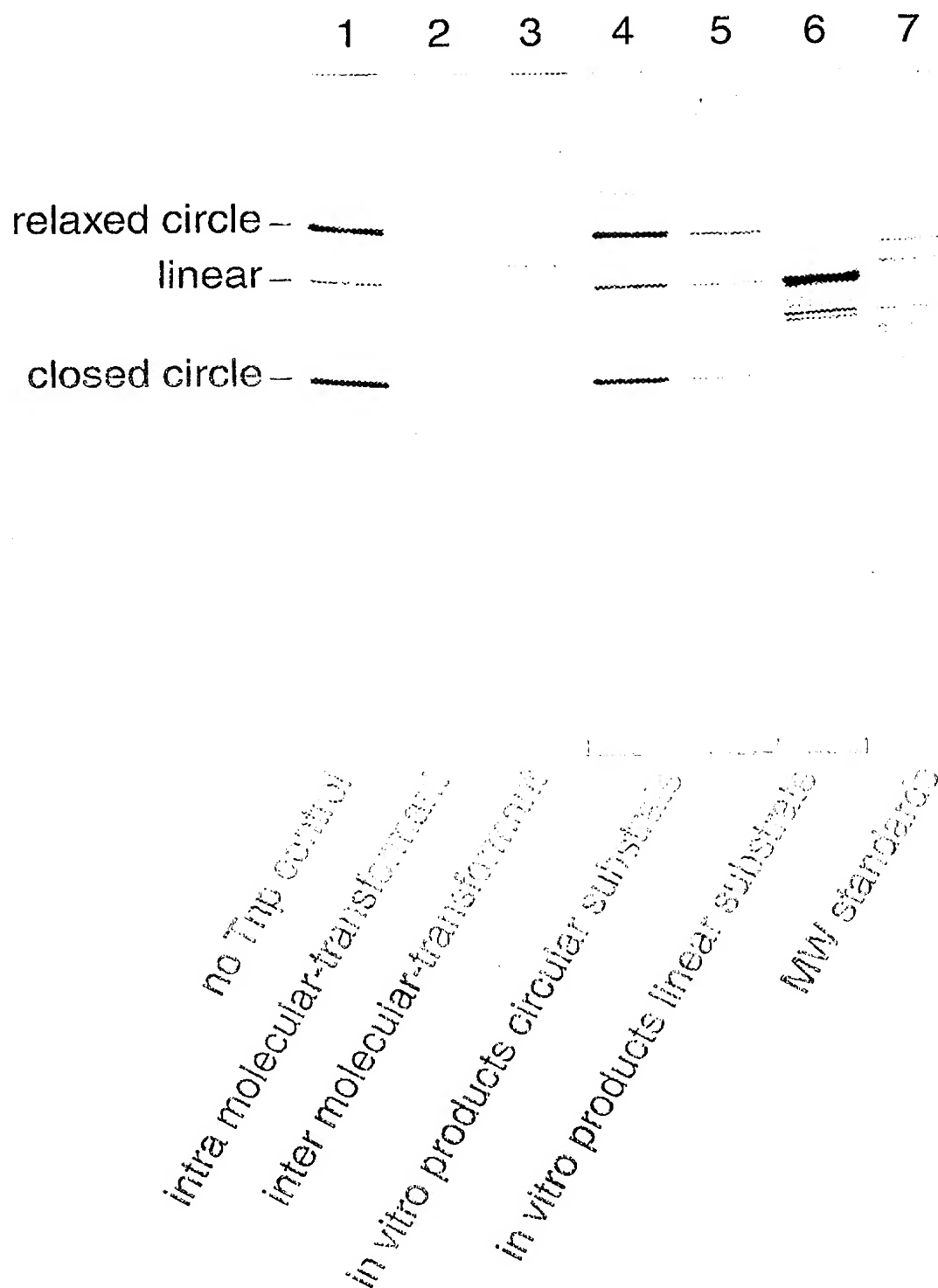


FIG 2
SUBSTITUTE SHEET (RULE 26)

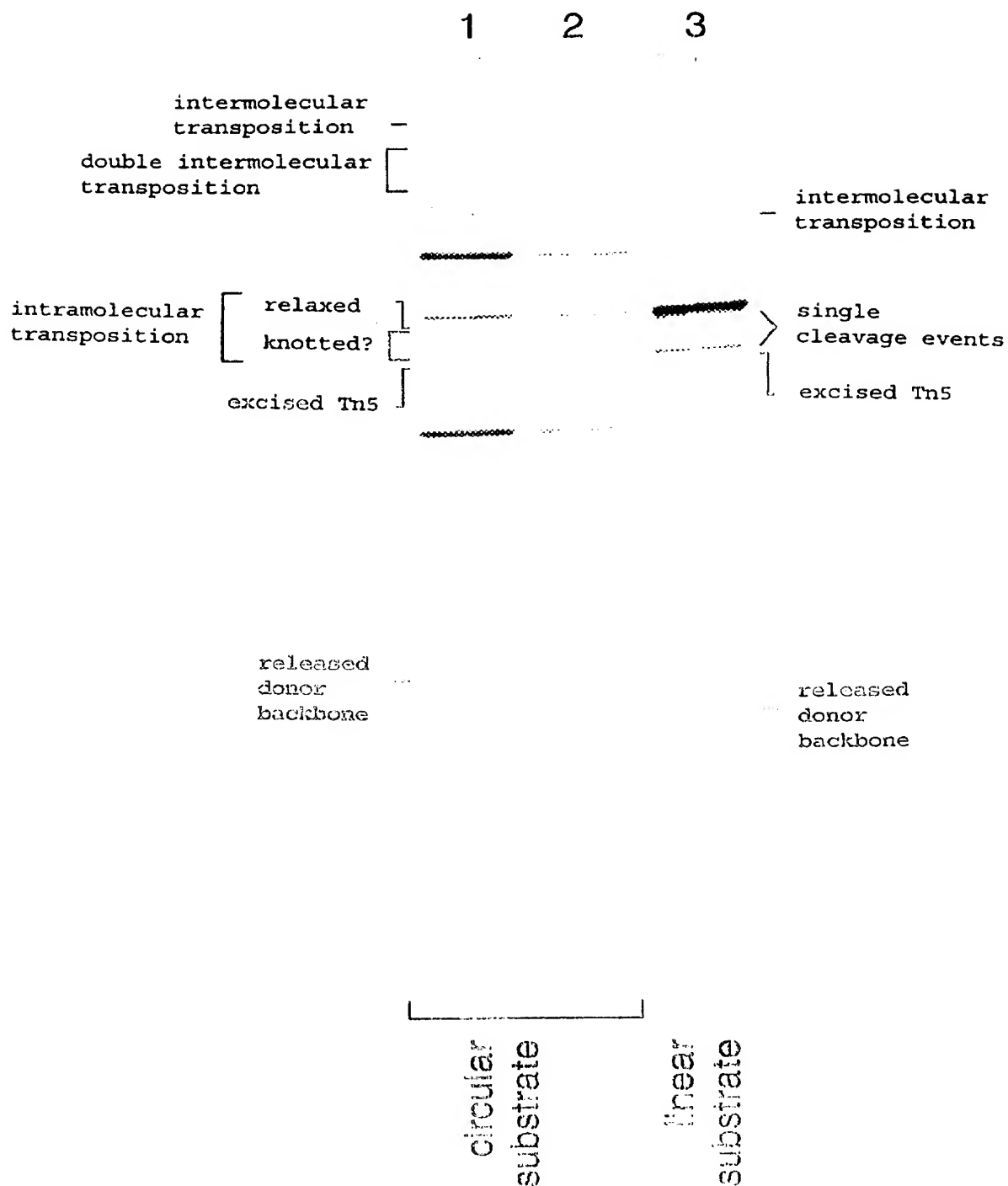


FIG 3
SUBSTITUTE SHEET (RULE 26)

4 / 8

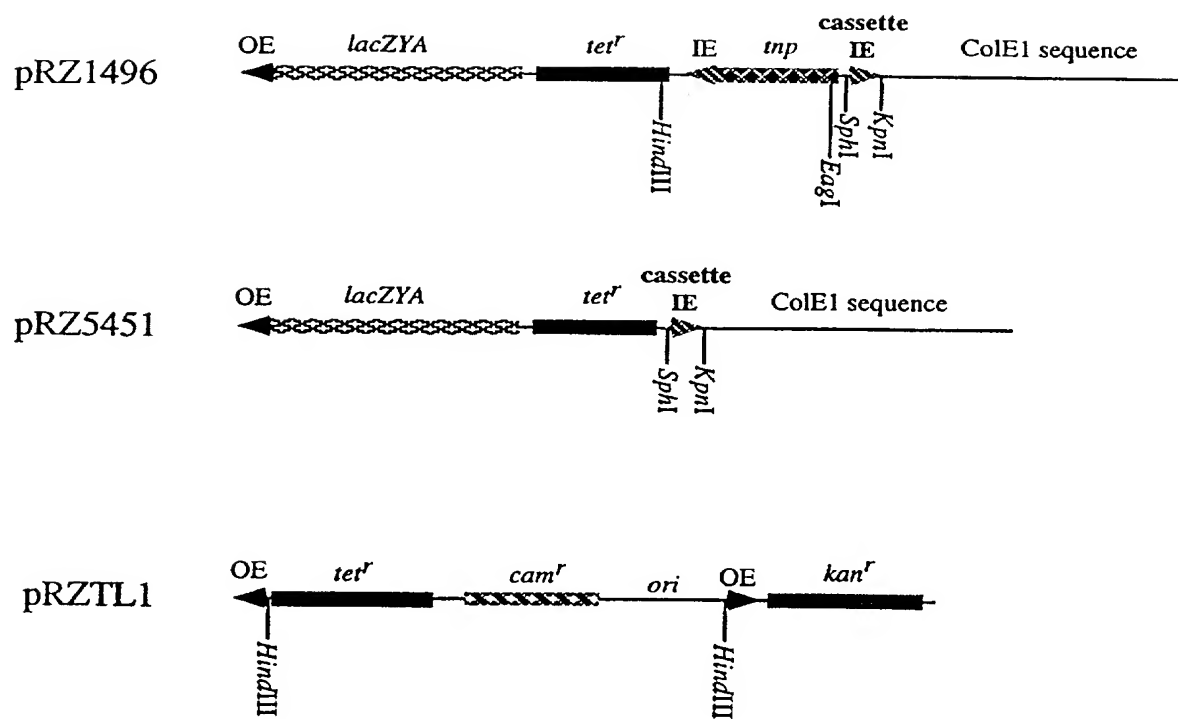
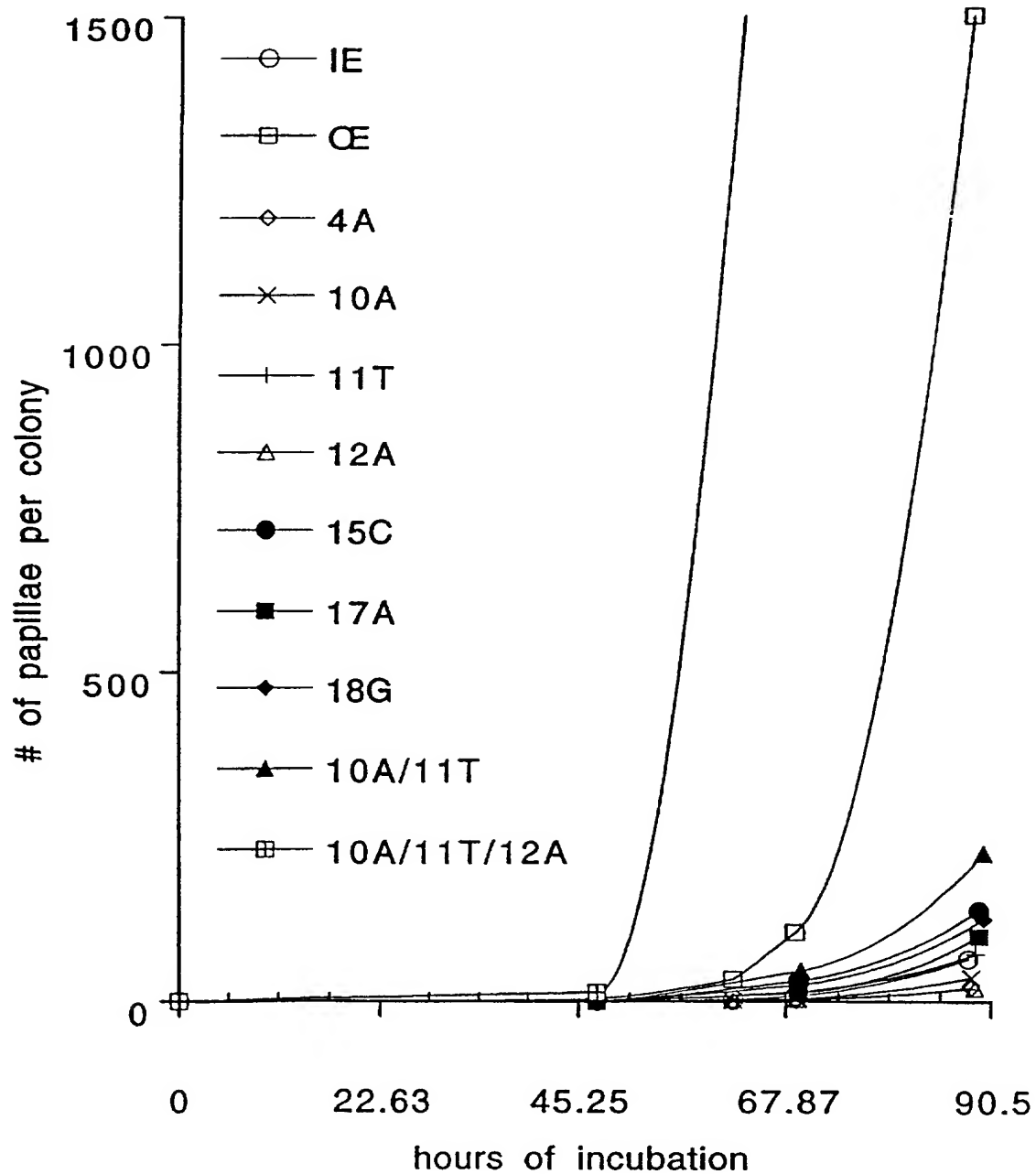


FIG 4

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a**Papillation of IE Mutants with EK54 Tnp****FIG 5**

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b

Papillation of IE Mutants with EK54 Tnp

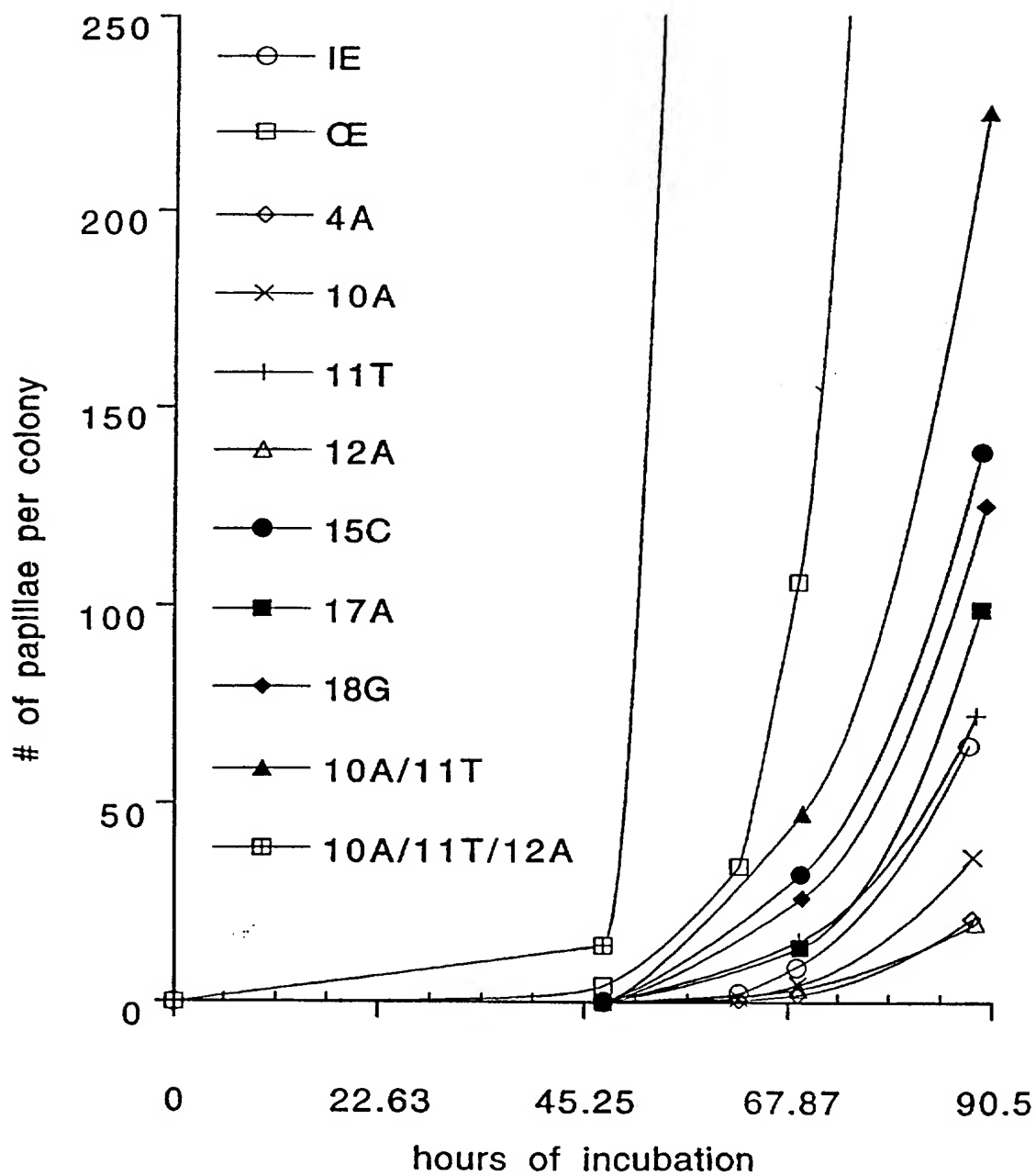
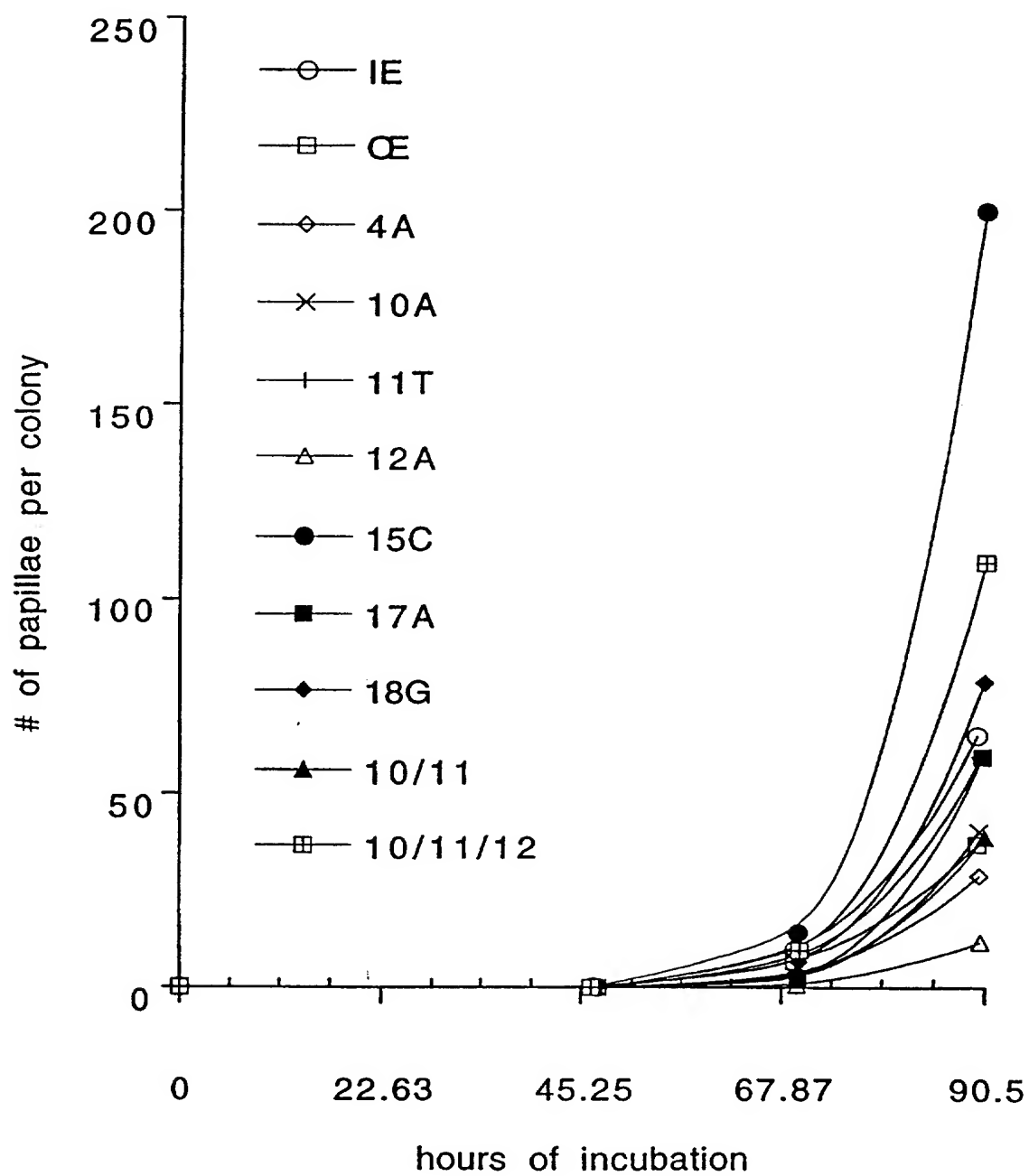
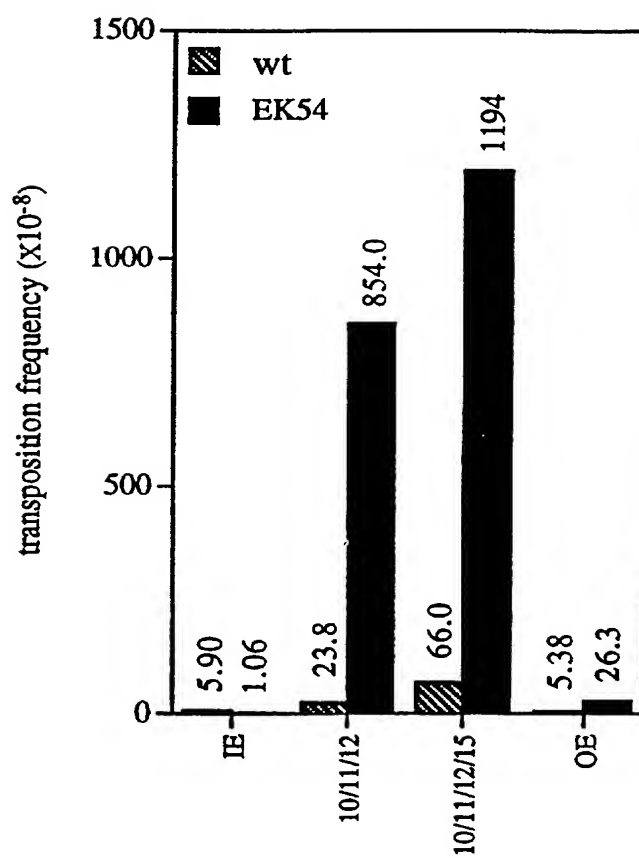


FIG 6

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Papillation of IE Mutants with wt Tnp**FIG 7**

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a**FIG 8**

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 97/15941

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/55 C12N9/22 C12N15/90 C12N15/85

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>ZHOU M ET AL: "Three types of novel mutations in the NH-2-terminus of Tn5 transposase: Structure-function of transposase."</p> <p>KEYSTONE SYMPOSIUM ON TRANSPOSITION AND SITE-SPECIFIC RECOMBINATION: MECHANISM AND BIOLOGY, PARK CITY, UTAH, USA, JANUARY 21-28, 1994. JOURNAL OF CELLULAR BIOCHEMISTRY SUPPLEMENT 0 (18B). 1994. 45. ISSN: 0733-1959, XP002052633 see the whole document</p> <p style="text-align: center;">--- -/--</p>	<p>1-3, 15-17, 22,23, 26,34-36</p>



Further documents are listed in the continuation of box C



Patent family members are listed in annex

* Special categories of cited documents :

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Date of the actual completion of the international search

20 January 1998

Date of mailing of the international search report

03/02/1998

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Hix, R

INTERNATIONAL SEARCH REPORT

International Application No

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WEINREICH M D ET AL: "Evidence that the cis preference of the Tn5 transposase is caused by nonproductive multimerization." GENES & DEVELOPMENT 8 (19). 1994. 2363-2374. ISSN: 0890-9369, XP002052634 cited in the application see the whole document ---	1,4,5, 15,18, 19,22, 24,26, 34,37,38
Y	DELONG, ALISON ET AL: "Trans-acting transposase mutant from Tn5" PROC. NATL. ACAD. SCI. U. S. A. (1991), 88(14), 6072-6 CODEN: PNASA6;ISSN: 0027-8424, 1991, XP002052635 see the whole document ---	1-47
Y	WIEGAND, TORSTEN W. ET AL: "Characterization of two hypertransposing Tn5 mutants" J. BACTERIOL. (1992), 174(4), 1229-39 CODEN: JOBAAY;ISSN: 0021-9193, February 1992, XP002052636 see the whole document ---	1-47
Y	WIEGAND, TORSTEN WALTER: "Transposase mutants that increase the transposition frequency of Tn5" (1993) 164 PP. AVAIL.: UNIV. MICROFILMS INT., ORDER NO. DA9315014 FROM: DISS. ABSTR. INT. B 1993, 54(6), 2886, 1993, XP002052637 see the whole document ---	1-47
Y	WEINREICH M D ET AL: "A functional analysis of the Tn5 transposase. Identification of domains required for DNA binding and multimerization." J. MOL. BIOL., vol. 241, 1993, pages 166-177, XP002052638 see the whole document ---	1-47
Y	JILK R A ET AL: "The organization of the outside end of transposon Tn5." JOURNAL OF BACTERIOLOGY, vol. 178, no. 6, March 1996, pages 1671-1679, XP002052640 see the whole document ---	1-47

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 97/15941

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	R.C. JOHNSON ET AL.: "DNA sequences at the ends of transposon Tn5 required for transposition." NATURE, vol. 304, 21 July 1983, pages 280-282, XP002052641 cited in the application see the whole document ---	1-47
P,Y	YORK, DONA ET AL: "Purification and biochemical analyses of a monomeric form of Tn5 transposase" NUCLEIC ACIDS RES. (1996), 24(19), 3790-3796 CODEN: NARHAD;ISSN: 0305-1048, 1996, XP002052642 see the whole document ---	1-47
P,X	ZHOU M ET AL: "Tn5 transposase mutants that alter DNA binding specificity." JOURNAL OF MOLECULAR BIOLOGY 271 (3). 1997. 362-373. ISSN: 0022-2836, XP002052643 see the whole document ---	1-47
P,X	YORK, DONA ET AL: "DNA binding and phasing analyses of Tn5 transposase and a monomeric variant" NUCLEIC ACIDS RES. (1997), 25(11), 2153-2160 CODEN: NARHAD;ISSN: 0305-1048, 1997, XP002052645 see the whole document -----	1-47